

Wound healing activity of flavonoid fraction isolated from the stem bark of *Butea monosperma* (Lam) in albino wistar rats

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ABSTRACT

The study aims to evaluate the wound healing properties of bioactive flavonoid fraction from the ethanolic extract of *Butea monosperma* (Lam) stem bark. In this study the wound healing activity of the flavonoid fraction isolated from the stem bark of *Butea monosperma* were evaluated in excision, incision and dead space wound healing models using Albino wistar rats. The wound healing activity was assessed by the breaking strength in case of incision wounds, epithelialization and wound contraction in case of excision wound and granulation tissue dry weight, breaking strength and hydroxyproline content in case of dead space wound. The flavonoid fraction showed the significant wound healing activity on all three wound models. The phytochemical investigations revealed that the flavonoid fraction contains two isoflavones genistein and prunetin. The increased rate of wound contraction and hydroxyproline content in the flavonoid fraction treated animals provides a scientific base to the ethno medicinal use of *Butea monosperma*, which is largely attributable to the additive or synergistic effect of isoflavones present in the flavonoid fraction.

Keywords: *Butea monosperma*, Flavonoid fraction, Dead space wound, Excision wound, Incision wound.

INTRODUCTION

Wound is a breach in the normal tissue continuum, resulting in a variety of cellular and molecular sequelae. The basic principles of optimal wound healing which include minimizing tissue damage, debriding nonviable tissue, maximizing tissue perfusion and oxygenation, proper nutrition and moist wound healing environment have been recognized for many years [1]. A number of drugs ranging from simple non-expensive analgesics to complex and expensive chemotherapeutic agents administered in the management of wound affect healing either positively or negatively [2]. Wounds are inescapable events of life which arise due to physical injury, chemical injury or microbial infections. Healing of wounds usually takes place in a direction away from its normal course and under healing, over healing or no healing of wounds is common. Management of under healing wounds is a complicated and expensive program and research on drugs that increase wound healing is a developing area in modern biomedical sciences. Several drugs obtained from plant sources are known to increase the healing of different types of wounds. Though some of these drugs have been screened scientifically for evaluation of wound healing activity in different pharmacological models and patients, the potential of many of the traditionally used herbal agents remain unexplored. In few cases active chemical constituents were identified [3].

Butea monosperma (Lam) (Fabaceae) is a medicinal plant growing in Burma, India and Sri Lanka. The flowers are tonic, astringent, aperient and diuretic. The decoction of the bark is traditionally used in cold, cough, fever, various forms of haemorrhages, in menstrual disorders and in the preparation of tonics and elixirs. The stem bark is reported to possess antitumour, antiulcer, antifungal and antidiarrhoeal activities [4-6]. It is also reported that the powder of the stem bark is used to apply on injury caused due to an axe, the juice of the stem is applied on goiter of human beings and the paste of the stem bark is applied in case of body swellings [7]. The roots are reported in the treatment of filariasis, night blindness, helminthiasis, piles, ulcers, and tumors [8]. It is reported that the ethanolic extract of seeds of *Butea monosperma*, on oral administration showed antifertility activity in mice and in rats [9]. Palsonin an active principle isolated from *Butea monosperma* seeds and its piperazine salt exhibited good anthelmintic activity *in vitro* on *Ascaris lumbricoides* and *in vivo* on *Taxicara canis* [8]. The petroleum ether extract and triterpene isolated from flowers of *Butea monosperma* exhibited anti convulsant activity [10, 11]. It has been reported that the methanolic extract of stem bark of *Butea monosperma* showed anti inflammatory and analgesic activity [12]. It has been reported that *Butea monosperma* stem bark extract and the flavonoid fraction isolated from it showed anti inflammatory activity [13, 14]. It is reported the efficacy of *Butea monosperma* on dermal wound healing in rats [15]. Recently we have reported the evaluation of wound healing properties of bioactive fractions from the extract of *Butea monosperma* (Lam) stem bark [16]. In continuation of the studies we have investigated the wound healing potential of flavonoid fraction isolated from the extract of *Butea monosperma* (Lam) stem bark and phytochemical investigations were carried out to isolate the active principles from *Butea monosperma* (Lam) stem bark.

MATERIALS AND METHODS

Plant material

The stem bark of *Butea monosperma* was collected during July 2009 from Manipal, Udupi district, Karnataka state, India. The samples were authenticated by Dr. Gopalakrishna Bhat, Professor of Botany, Poorna Prajna College, Udupi, India. A herbarium specimen has been deposited at the college for further reference.

Preparation of plant extracts

The bark was dried in the shed and coarsely powdered. The powder was extracted with ethanol in a soxhlet apparatus for 72h. The ethanolic extract was evaporated in vacuo giving the residue (24%). The ethanolic extract obtained was suspended in distilled water in small amounts and was extracted successively and exhaustively with petroleum ether (60-80°C), benzene, chloroform and acetone in the order of increasing polarity. The extract and fractions were concentrated in a rotary evaporator at reduced pressure.

Preliminary phytochemical analysis

In our previous studies the acetone fraction of the ethanolic extract was found to be having the biological activity [16]. Hence the bioactive acetone fraction was subjected to phytochemical screening according to the phytochemical methods described by Harborne [17].

Chromatographic separation and isolation of constituents:

The acetone fraction was chromatographed by column chromatography over silica gel and eluted with n-hexane/ethyl acetate (80:20 v/v) with increasing amounts of ethyl acetate. 10 mL of the eluates were collected and monitored with TLC and the similar fractions were combined together. Total 4 fractions were collected and the fraction 3 was identified as flavonoid fraction. The bioactive flavonoid fraction was further separated by HPLC (ODS, 250X10 mm, 5μ) using ethyl acetate/methanol/formic acid (70:30:3 v/v/v) as mobile phase to afford compounds **1** and **2**. The structure elucidation of compound **1** and **2** were done by comparing with reference data previously reported from available literature [18-22], and by co-TLC with the authentic samples.

Experimental animals

Adult Wistar strain rats (150 to 200 gm) were used for all the experiments in the present study. The animals were maintained under standard husbandry conditions in the animal house of the institute (temperature $25 \pm 2^{\circ}\text{C}$) in a natural light-dark cycle and fed with standard rodent diet and water *ad libitum*. Ethical committee clearance was obtained from IAE (Institutional Animal Ethics Committee) of CPCSEA (Ref. No./IAEC/XII/08/CLBMCP/2009-2010).

Acute toxicity studies

The acute toxicity of flavonoid fraction of *Butea monosperma* stem bark was determined as per the OECD guideline no. 423 (Acute toxic class method) [23]. Based on the results obtained from this study, the dose for wound healing activity was fixed to be 25 mg kg^{-1} b.w. and 50 mg kg^{-1} b.w. for dose dependent study.

Excision wound model

The rats were inflicted with excision wounds as described by Morton and Malone [24]. An excision wound was inflicted by cutting away 500 mm² full thickness of a pre-determined area on the depilated back of the rat. The rats were divided into four groups of six animals each. Group 1(control) animals were topically applied with simple ointment base, Group 2 animals were topically applied with soframycin ointment and the remaining groups were topically treated with 25 mg kg⁻¹ b.w. and 50 mg kg⁻¹ b.w. of test substances mixed with ointment base. Treatments were given once daily till the wound was completely healed. Epithelialization period was noted as the number of days after wounding required for the dead tissue remnants to fall off leaving no raw wound behind. Wound contraction rate was monitored by planimetric measurement of the wound area on alternate days. This was achieved by tracing the wound on a graph paper. Reduction in the wound area was expressed as percentage of the original wound size [25].

Incision wound model

The method of Ehrlich and Hunt was adopted for incision wound study [26]. The animals were anaesthetized under light ether, on the depilated backs of the animals, two paravertebral incisions of 6 cm length were made cutting through the full thickness of the skin. Interrupted sutures, 1 cm apart, were placed to approximate the cut edges of the skin. The rats were divided into four groups of six animals each. Group 1(control) animals were topically applied with simple ointment base, Group 2 animals were topically applied with soframycin ointment and the remaining groups were topically treated with 25 mg kg⁻¹ b.w. and 50 mg kg⁻¹ b.w. of test substances mixed with ointment base. The sutures were removed on the 8th post wound day and skin breaking strength was measured on the 10th day by continuous water flow technique [27].

Dead space wound model

Dead space wounds were created through a small transverse incision made in the lumbar region. A polypropylene tube (2.5 × 0.5 cm) was implanted subcutaneously beneath the dorsal paravertebral lumbar skin [28]. The day of the wound creation was considered as day zero. The animals were divided in to three groups of six animals each. Group 1 was the control group that received 2 mL of 1% carboxymethyl cellulose. The remaining groups were administered each with 25 mg kg⁻¹ b.w. and 50 mg kg⁻¹ b.w. of test substances orally, once daily for 10 days. Granulation tissue formed on the polypropylene tube was harvested by careful dissection on day 10 and the breaking strength of the granulation tissue was measured. The granulation tissue was dried in an oven at 60°C overnight and the dry weight was noted. Acid hydrosylate of the dry tissue was used for the determination of the hydroxyproline content [29].

Statistical analysis

The experimental results were expressed as mean ± S.E.M. Results were analyzed by the one- way ANOVA followed by Tukey-kramer post hoc multiple comparison test using Graph pad InStat version 3.00. P value of <0.05 was considered as statistically significant.

RESULTS

Preliminary phytochemical analysis

The preliminary phytochemical analysis of flavonoid fraction showed the presence of flavonoids, phenolic compounds, and steroids. From the flavonoid fraction we isolated two isoflavones where the compound 1 was identified as Genistein (Figure 1: 4^r, 5, 7-trihydroxy isoflavone) and the compound 2 (Figure 2: 4^r, 5-dihydroxy, 7-methoxy isoflavone) was identified as prunetin.

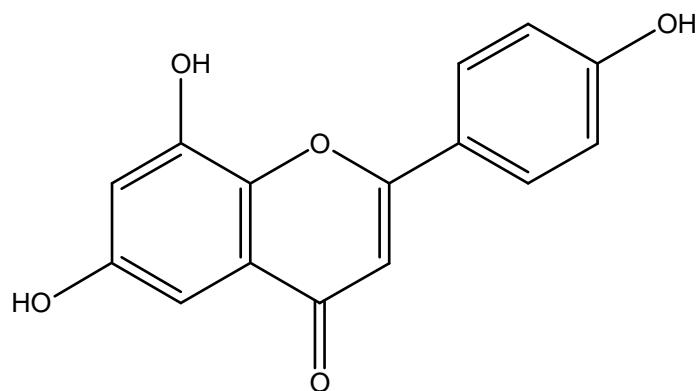


Figure 1: The structure of Genistein

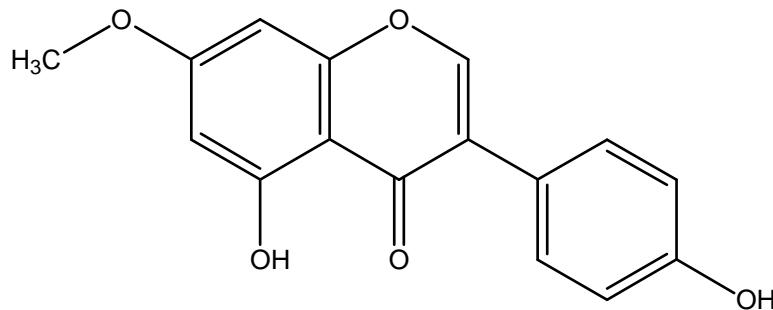


Figure 2: The structure of Prunetine

Excision wound model

Excision wounds heal by contraction (wound closure) and epithelialization, the percentage of wound closure or closure rate includes by recording the changes in wound area at fixed intervals of time, viz. 4th, 8th, 12th and 16th day after treatment with flavonoid fraction. The percentage of wound contraction in all the test groups was significant statistically when compared to control on the 4th day, there was no statistical significance in the percentage of wound contraction during the 8th and 12th days. But on 16th day there was a statistically significant difference in percentage of wound contraction. The maximum percentage of wound closure on the 16th day was observed with standard drug, soframycin (97.67%) and the flavonoid fraction at the test doses of 25 mg kg⁻¹ b.w. and 50 mg kg⁻¹ b.w. showed percentage wound closure of 90.17% and 95.50% respectively. The flavonoid fraction was also significantly (P < 0.05) reduced the epithelialization period of excision wounds. (Table 1).

Incision wound model

Incision wounds heal by granulation and collagenation. The mean wound breaking strength or tensile strength of wound in control group was 150.50g, while in the case of flavonoid fraction at the test doses of 25 mg kg⁻¹ and 50 mg kg⁻¹ showed 172.50g and 282.83g of mean breaking strength respectively. It was found that the mean time for epithelialization and mean scar area were reduced significantly, thereby increasing the mean tensile strength compared to control group. The mean wound breaking strength in case of soframycin treated group was 342.17 g (Table 2).

Dead space wound model

The mean dry weight of granulation tissue in the control group was 45.17mg, which was significantly (P < 0.05) increased to 59.17mg and 73.83mg in flavonoid fraction treated groups at test doses of 25 mg kg⁻¹ and 50 mg kg⁻¹ respectively when compared to control group. The breaking strength in control group was 330.67g, whereas the flavonoid fraction at the test doses of 25 mg kg⁻¹ (406.33g) and 50 mg kg⁻¹ (438.50g) showed significant increase in breaking strength when compared to the control group. The hydroxyproline content was significantly increased (P < 0.05) in flavonoid fraction treated groups at both test doses when compared to the control group (Table 3).

Table 1: Effect of flavonoid fraction of *Butea monosperma* stem bark on percentage of wound contraction and period of epithelialization in excision wound model

Groups	% Wound contraction				Period of epithelialization (days)
	4 th day	8 th day	12 th day	16 th day	
Control	5.98±0.157	59.0±0.36	84.67±0.8	85.33±0.71	23.33±0.80
Standard	65.0±1.57**	60.17±1.4	86.17±0.91	97.67±0.42**	15.17±0.31**
FLAV25	44.67±1.31**	59.17±0.48	85.67±1.02	90.17±0.48**	19.17±0.31**
FLAV50	60.17±0.6**	59.67±1.08	86.50±0.56	95.50±0.56**	16.17±0.48**

Standard: Soframycin ointment, FLAV25: Flavonoid fraction at dose 25 mg kg⁻¹ b.w. FLAV50: Flavonoid fraction at dose 50 mg kg⁻¹ b.w. Each value is the Mean ± S.E.M for 6 rats. **P<0.001 compared with control

Table 2: Effect of flavonoid fraction of *Butea monosperma* stem bark on wound breaking strength in incision wound model

Groups	Mean wound breaking strength (g)
Control	150.50±2.03
Standard	342.17±4.64**
FLAV25	172.50±3.86*
FLAV50	282.83±3.48**

Standard: Soframycin ointment, FLAV25: Flavonoid fraction at dose 25 mg kg⁻¹ b.w. FLAV50: Flavonoid fraction at dose 50 mg kg⁻¹ b.w. Each value is the Mean ± S.E.M for 6 rats. *P < 0.01; **P < 0.001 compared with control

Table 3: Effect of flavonoid fraction of *Butea monosperma* stem bark on dry weight, tensile strength and hydroxyproline content in dead space wound model

Groups	Dry weight (mg)	Tensile strength (g)	Hydroxyproline (µg/mg)
Control	45.17±0.48	330.67±3.11	22.01±0.14
FLAV25	59.17±1.45**	406.33±5.2**	23.55±0.19**
FLAV50	73.83±0.75**	438.50±4.64**	26.60±0.21**

FLAV25: Flavonoid fraction at dose 25 mg kg⁻¹ b.w. FLAV50: Flavonoid fraction at dose 50 mg kg⁻¹ b.w. Each value is the Mean ± S.E.M for 6 rats. **P<0.001 compared with control

DISCUSSION

The results of the present investigations revealed that the flavonoid fraction isolated from the stem bark of *Butea monosperma* possess significant wound healing activity in excision, incision and dead space wound models. In spite of tremendous development in the field of synthetic drugs during recent era, they are found to have some or other side effects, whereas plants still hold their own unique place, by the way of having no side effects. Therefore, a systematic approach should be made to find out the efficacy of plants against wounds so as to exploit them as herbal wound healing agents.

Experimental assessment of the wound healing activity of flavonoid fraction showed increased rate of wound contraction and epithelialization and increased granuloma tissue formation. Topical application of the flavonoid fraction on excision wounds accelerated wound contraction and reduced epithelialization period in rats. Wound healing involves regeneration of specialized cells by proliferation of surviving cells and connective tissue response characterized by the formation of granulation tissue [30]. It is also characterized by haemostasis, reepithelialization and remodeling of the extracellular matrix. Epithelialization, which is the process of epithelial renewal after injury, involves the proliferation and migration of epithelial cells towards the centre of the wound while wound contraction is largely due to the action of myofibroblasts [31, 32]. Thus, the effect of flavonoid fraction on wound contraction and epithelialization suggest it may enhance epithelial cells migration and proliferation, as well as the formation, migration and action of myofibroblasts. On chronic oral administration, flavonoid fraction enhanced the granuloma tissue formation in dead space wounds. Granuloma tissue formed on an inert foreign body in a dead space comprises an accumulation of modified macrophages [30], histological giant cells and undifferentiated connective tissue consisting largely of collagen [32, 33]. Increase in granuloma tissue in dead space wound is associated with enhanced collagen maturation and increased protein content as well as angiogenesis [34-36] in the wound. These processes are indicators of new tissues generation and suggest that the flavonoid fraction may stimulate mechanisms associated with tissue regeneration. Closely related to this is the effect of growth factors secreted by macrophages on wounds. Macrophages secrete peptide growth factors that exert pro-healing effect by stimulating regeneration, fibroblast proliferation and activation and angiogenesis [30]. It is, therefore, likely that in addition to enhancing wound contraction and epithelialization, the flavonoid fraction may also stimulate processes associated with tissue regeneration.

The flavonoid fraction significantly increased the skin breaking strength and hydroxyproline content which was a reflection of increased collagen levels by increased cross linking of collagen fibres. In addition, increase in dry granulation tissue weight indicated the presence of higher protein content [37]. The breakdown of collagen liberates free hydroxyproline and its peptides and elevated level of hydroxyproline is the index of increased collagen turnover.

From the above studies it is quite apparent that the flavonoid fraction of *Butea monosperma* stem bark possesses significant wound healing activity, which was evident by the increased rate of wound contraction, reduction in the period of epithelialization, increase in collagen deposition, breaking strength and hydroxyproline in granulation tissue.

CONCLUSION

The flavonoid fraction of *Butea monosperma* stem bark showed significant wound healing property which provides a scientific base to the ethno medicinal use of *Butea monosperma*. We propose that the presence of some bioactive compounds such as the isoflavones genistein and prunetin in the flavonoid fraction of *Butea monosperma* were responsible for its potent wound healing property. The present investigation offers scientific evidence to the folkloric accounts of the use of stem bark in treating cuts and wounds.

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REFERENCES

[1] G.F. Pierce, T.A. Mustoe, Pharmacologic enhancement of wound healing, *Annu Rev Med*, **1995**, 46, 467–481.

[2] D. Prasad, C.M. Rao, *Ind J Exp Biol*, **1995**, 33, 845–847.

[3] T.K. Biswas and B. Mukherjee, *Int J Low Extrem Wounds*, **2003**, 2, 25-39.

[4] B.M.R. Bandara, N.S. Kumar, K.M.S. Wimalasiri, *J.Ethanopharmacol*, **1989**, 25, 73.

[5] B.M.R. Bandara, N.S. Kumar, K.M.S. Wimalasiri, *J. Natl. Sci. Comu (Sri Lanka)*, **1990**, 18, 97.

[6] A. Gunankunru, K. Padmanaban, P. Thirumal, J. Pritila, G. Parimala, N. Vengtesan et al. *J. Ethanopharmacol*, **2005**, 98, 241-244.

[7] D.A. Patil, P. Shubhangi, M.V. Patil, *Natural Product Radiance*, **2006**, 5(4), 323-325.

[8] R.K. Raj, P.A. Kurup, *Indian J Med Res*, **1968**, 56(12), 1818-25.

[9] M.K. Razdan, K. Kapila, N.K. Bhide, *Indian J Physiol Pharmacol*, **1969**, 13(4), 239-49.

[10] V.S. Kasture, C.T. Chopde, V.K. Deshmukh, *J Ethnopharmacol*, **2000**, 71(1-2), 65-75.

[11] V.S. Kasture, V.K. Deshmukh, C.T. Chopde, *Phytother Res*, **2002**, 16(5), 455-60.

[12] M.W. Carey, G. Krishna Mohan, *Pharmacologyonline*, **2007**, 2, 88-94.

[13] A. Muralidhar, K. Sudhakar Babu, T. Ravi Sankar, P. Reddanna, G.V. Reddy, J. Latha, *International Journal of Phytopharmacology*, **2010**, 1(2), 124-132.

[14] A. Muralidhar, K. Sudhakar Babu, T. Ravi Shankar, P. Reddanna, G.V. Reddy, J. Latha, *International Journal of Pharmacy & Therapeutics*, **2010**, 1(2), 44-51.

[15] M. Sumitra, P. Manikandan, L. Suguna, *Journal of Biochemistry and Cell Biology*, **2005**, 37, 566–573.

[16] A. Muralidhar, K. Sudhakar Babu, T. Ravi sankar, P. Reddanna, J. Latha, *International Journal of Phytomedicine*, **2011**, 3, 41-49.

[17] J.B. Harborne, *Phytochemical Methods: A guide to Modern Techniques of Plant Analysis*, 2nd edition, Chapman and Hall, London, **1998**, 282.

[18] J.B. Harborne, In *The Flavonoids: Advances in Research since 1986*; Chapman & Hall: London, U.K., 1994, 458.

[19] P.K. Agrawal, In *Carbon-13 NMR of Flavonoids*; Ed.; Elsevier Science Publishers: Amsterdam, **1989**; pp 102, 196.

[20] G.N. Muraleedharan, W. Haibo, M.S. Gale, M.B. Alden, J. Ian Gray, *J. Agric. Food Chem.*, **1999**, 47, 840-844.

[21] A.C. Talukdar, N. Jain, S. De, H.G. Krishnamurthy, *Phytochemistry*, **2000**, 53, 155-157.

[22] P. Alexandra, F.V. Rute, M.F. Helder, T. Generosa, B. Carlos, R. Ana Isabel, T. Adriano, *Journal of Ethnopharmacology*, **2004**, 93, 363–370.

[23] OECD, 2002, Acute oral toxicity, Acute oral toxic class method guideline 423 adopted 23.03.1996. In: Eleventh Addendum to the OECD guidelines for the testing of chemicals organisation for economical co-operation and development. Paris. June **2000**.

[24] J.J.P. Morton, M.H. Malone, *Arch Int Pharmacodyn*, **1972**, 196, 117-126.

[25] S.B. Nayak, S.S. Raju, M. Eversley, A. Ramsubhag, *Phytother Res*, **2009**, 23, 241-5.

[26] H.P. Ehrlich, T.K. Hunt, *Ann Surg*, **1969**, 170(2), 203–206.

[27] K.H. Lee, *J Pharma Sci*, **1968**, 57(7), 1238–1240.

[28] S. Nayak, S.G. Rao, K.D. Murthy, S.N. Somayaji, K.L. Bairy, *Indian J Expt Biol*, **2003**, 41(6), 645–648.

[29] R.E. Neuman, M.A. Logan, *J Biol Chem*, **1950**, 184, 299.

[30] K. Whaley, A.D. Burt, Inflammation, Healing and Repair. In: Muir's Textbook of Pathology, MacSween, R.M.N. and K. Whaley (Eds.). 13th Edn., Arnold, London ISBN: 0340569573, **1996**, 112-165.

[31] R.S. Cotran, V. Kumar, S.L. Robbins, F.J. Schoen, Inflammation and Repair. In: Robbins Pathologic Basis of Disease, 5th Edn, W.B. Saunders Company, Pennsylvania, ISBN: 0-7216-5032-5, **1994**, 51-92.

[32] H. Mohan, Inflammation and Healing, In: Textbook of Pathology, 5th Edn., Jaypee Brothers, New Delhi, ISBN: 81-8061-368-2, **2005**, 133-179.

[33] K.L. Bairy, C.M. Rao, *J Natural Remedies*, **2001**, 1, 25–27.

[34] S. Azad, Essentials of Surgery, Paras Medical Publications, Hyderabad, ISBN: 8181911180, **2002**, 1.

[35] K.H. Swamy, V. Krishna, K. Shankarmurthy, A.B. Rahiman, K.L. Mankani, K.M. Mahadevan, *J Ethnopharmacol*, **2007**, 109, 529-34.

[36] B.G. Harish, V. Krishna, H.S. Kumar, K.B. Ahamed, R. Sharath, K.H. Swamy, *Phytomedicine*, **2008**, 15, 763-767.

[37] B.K. Manjunatha, *Indian Drugs*, **2006**, 43, 835.