

Research Journal of Oncology

Open access Research article

Vimentin and Ki-67 Acting as Immunotherapy Predictive Biomarkers in Pulmonary Carcinomas Transthoracic and Bronchial Biopsies

Maria Inês Figueiredo^{1,2*}, Ana Ladeirinha², Ana Alarcão², Vítor Sousa^{1,2,3}, Lina Carvalho^{1,2,3}

ABSTRACT

Introduction: Programmed death-ligand 1 (PD-L1) expression became a routine biomarker to preview response to programmed death-1 (PD-1)/PD-L1 inhibitors, with diverging parameters concerning PD-L1 scoring and variable response to immunotherapy agents. The aim of this study was to evaluate association between PD-L1 expression and immunohistochemistry panel applied in Pathology practice, defining any of those antibodies as biomarkers concurrent in patient's selection for PD-1/PD-L1 blockade therapy.

Methods: A total of 97 cTNM IIIb/IV staged pulmonary carcinoma biopsies were randomly selected between 2018/2020, after adequate representativeness and PD-L1 expression scored through Dako 22C3 antibody. The panel with cytokeratin 7, thyroid transcription factor 1 (TTF1), cytokeratin 5.6, cluster of differentiation 56 (CD56), periodic acid-Schiff (PAS-D), vimentin expression, and ki-67 labeling index (LI) was considered for retrieving reports and respective archival slides.

Results: PD-L1 expression in tumor cells (TCs) was identified in 56 samples and significantly associated with male gender (p=0.028), vimentin expression (p=0.018) and ki-67 LI>30% (p=0.029). A tendency to PD-L1 positivity came up in lymphocytic-predominant/immune-inflamed stroma (9/10), adenocarcinoma solid subtype (21/23) and CK7-negative squamous cell carcinomas (8/13). When more than 50% TCs expressed PD-L1, the risk of vimentin expression was 3.85 times higher (OR=3.85; p=0.013), and for ki-67 LI>30% the risk was 9.90 times higher (OR=9.90; p=0.033), compared with PD-L1-negative samples.

Conclusion: High proliferation status defined by ki-67 LI>30% and epithelial-mesenchymal transition phenotype verified by vimentin staining analysis might complement PD-L1-positive TCs percentage determination for immunotherapy prescription. These patients will more likely benefit from PD-1/PD-L1 blockade therapy, overcoming the limitations of patient selection based on PD-L1 immunohistochemistry status.

Keywords: Pulmonary Carcinoma; PD-L1, Immunotherapy; Epithelial-Mesenchymal Transition

INTRODUCTION

Lung cancer remains clinically asymptomatic in early stages and 75% of cases are diagnosed at advanced stages, out of surgical staging resection and with 5-year survival rate of approximately 15% [1-3]. Targeted therapies with tyrosine kinase

inhibitors (TKIs) have become the standard of care to approximately 20% of patients with pulmonary carcinomas, for the last two decades [3,4].

Programmed death-1 (PD-1)/programmed death-ligand 1 (PD-L1) inhibitors are currently applied in combination with pemetrexed and carboplatin as first-line therapy in adenocarcinomas

Received: 25-January-2022 Manuscript No: iprjo-22-12390 Editor assigned: 27- January -2022 **PreQC No:** iprjo-22-12390 (PQ) Reviewed: 10-February-2022 QC No: iprjo-22-12390 **Revised:** 15-February-2022 Manuscript No: iprjo-22-12390 (R) **Published:** 22- February -2022 DOI: 10.36648/iprjo.6.2.5

Corresponding author Maria Inês Figueiredo, Department of Medicine, University of Coimbra, Portugal, E-mail: ines_500@ hotmail.com

Citation Maria IF, Vítor S, Lina C, Ana L, Ana A (2022) Vimentin And Ki-67 Acting As Immunotherapy Predictive Biomarkers In Pulmonary Carcinomas Transthoracic And Bronchial Biopsies. Res J Onco Vol.6:2

Copyright © Maria IF, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

¹Department of Medicine, University of Coimbra, Portugal

²Department of Anatomical and Molecular Pathology, University of Coimbra, Portugal

³Department of Anatomical Pathology, University Hospital of Coimbra, Portugal

(ADCs), regardless of PD-L1 expression [5]. For pembrolizumab, however, PD-L1 expression determined by 22C3 Dako stain is still mandatory for first-line therapy upon over 50% expression in tumor cells (TCs).

As a continuous biomarker within tumoral heterogenous compartments, without standardization among different assays, PD-L1 antibodies have been referred for the available drugs, after different detection antibodies and scoring systems [3,6]. Blueprint Comparison Project demonstrated high concordance among 3 of the 4 currently used assays, with the limitation of including 39 tumor surgical samples [7], and Blueprint Phase 2 corroborated these results in 81 surgical samples [8].

While high PD-L1 expression levels correlated with increased response to PD axis blockade therapy in several studies [3], some carcinomas harboring PD-L1-positive cells did not respond to immunotherapy, and 10%-20% of responses to anti-PD therapy occurred in PD-L1-negative tumor biopsies [9-11]. Hence, since PD-L1 expression may not be an efficient predictive biomarker of response, but rather a risk factor used to select patients more likely to benefit from immunotherapy, additional predictive cost-effective biomarkers are needed to identify potential responders to immunotherapy and be considered in follow-up [12].

To overcome the limitations of PD-L1 immunohistochemistry (IHC) expression determination, Tumor mutation burden (TMB), evaluated by next-generation sequencing (NGS) [6], is emerging with predictive score for response to immunotherapy [10,13,14]. Rizvi et al. demonstrated that progression free survival (PFS) and clinical response to PD-L1 inhibitors were higher in patients with high TMB tumors, irrespective of PD-L1 status [15]. However, major drawbacks arise regarding TMB evaluation: it is not yet routinely used in clinical practice owing to elevated costs and interpretation complexity due to low versus high levels scores variability [10,13].

Contrary to the emerging TMB, IHC is definitely considered fast and cost-effective in routine Pathology practice, aiding the identification of predictive biomarkers of response to lung cancer therapies [16]. Routine biopsies morphology and IHC panels support diagnosis, classification and screening for therapeutical targets in pulmonary carcinomas [16]. Defining tumoral histopathology subtyping with final diagnosis based on routine IHC panels allows: basic classification among incompletely represented tumors in biopsy samples, minimization of diagnostic mistakes, and exclusion of metastatic origin and selection of samples for molecular testing and therapy guidance [17]. Recognition of ADCs patterns, namely solid, papillary, micropapillary, acinar and mucinous differentiation, and keratinizing versus non-keratinizing squamous cell carcinomas (SQCs), became feasible in Pathology practice [17].

A consistent panel of IHC antibodies, such as thyroid transcription factor 1 (TTF1) and NapsinA (both expressed in more than 85% of lung ADCs), cytokeratin (CK) 5/6 and p40 (to establish squamous cell differentiation), vimentin (mesenchymal marker) and proliferation marker ki-67 labeling index (LI), will change over 90% of biopsies sampling to correctly classify ADCs and SQCs, including other mixed subtypes [16].

The aim of this study was to evaluate the association between routine IHC panel applied to bronchial and pulmonary biopsies for pulmonary carcinomas classification, according to World Health Organization (WHO) 2015/2020 definitions, and PD-

L1 status (22C3 Dako antibody). Proliferation marker ki-67 LI, dedifferentiation marker vimentin and tumoral stroma characteristics in association with PD-L1 expression were also considered in this study, to search patient selection for PD-1/PD-L1 blockade therapy in pulmonary carcinomas based in archival biopsy tissue, integrating the rationale for selecting the cases.

MATERIALS AND METHODS

Tumor samples

Based on biopsy diagnosis of non-surgical stages bronchopulmonary carcinomas, cT3b and cT4 by 2017 TNM system, a series of 97 cases concerning 16 SQCs, 64 ADCs, 7 adenosquamous carcinomas (ADSQCs), 3 carcinomas not otherwise specified (NOS) and 7 pleomorphic carcinomas were included in this study. The cases were selected according to representative tumor tissue: at least three fragments in bronchial biopsies and similar area either in transthoracic and pleural biopsies, also represented in fragmented small surgical biopsies and pleural biopsies, consecutively collected in 2018/2020; neuroendocrine pattern/IHC (cluster of differentiation 56 (CD56)) expression were exclusive. WHO 2015/2020 classification for lung tumors, currently applied to biopsy specimens at the University Hospital of Coimbra, was well defined in this series of samples, according with representative tumor tissue, as referred.

ADC subtyping was registered according to the represented predominant pattern, as solid (23 cases), mucinous (22 cases) and acinar (12 cases); micropapillary designation was prevalent when this pattern was present (7 cases).

Median age of diagnosis was 68 years, ranging from 43 to 96 years; 75 patients were male and 22 were female. Descriptive data is summarized in **Table 1**. The study fulfilled the rules for archival retrospective study defined by the Faculty of Medicine of the University of Coimbra Ethical Committee.

Immunohistochemistry

In order to ascertain tumor subgroups, CK7, TTF1, CK5/6, CD56, ki-67 LI and vimentin antibodies had been applied according to routine protocols. Periodic acid-Schiff (PAS-D) staining was performed in all carcinomas following the McManus Technique with diastase for glycogen digestion.

Concerning PD-L1, formalin-fixed paraffin-embedded (FFPE) serial sections of 3 μm were mounted on positively charged slides, deparaffinized and stained for PD-L1 using Food and Drug Administration (FDA)-approved Dako PD-L1 22C3 antibody (Dako, Carpinteria, CA). On Ventana platform (LDT), sections were incubated in 3% diluted hydrogen peroxide for 5 minutes to neutralize endogenous peroxidase activity. Non-specific binding of primary antibodies and polymer were reduced with Protein Block. 22C3 Dako antibody, at 1:40 dilution, had been applied to tumor sections and then incubated for 52 minutes. After tris-buffered saline (TBS) washing, Post Primary Block was used to enhance penetration of anti-mouse/ rabbit IgG HRP-polymer; 3,3'-diaminobenzidine (DAB) was used as chromogen. Finally, 0.02% diluted hematoxylin was used to counterstain the sections. Positive and negative controls were used, with human tonsil tissue as positive control for the PD-L1 staining (Supplementary Table 1).

The applied IHC panel described in **Supplementary Table 1**, followed manufacturer indications. The slides were evaluated and

scored in light microscopy by two experienced pathologists.

IHC scoring

In general, 50% cut-off was defined for the applied routine antibodies, considering 3+ as high positivity. Positivity was near 100% for CK5/6 in SQCs and for CK7/TTF1 duet in ADCs. Vimentin expression cut-off was established also at 50% when expressed in TCs, as well as for PAS-D-positive cells interpretation, allowing two groups definition. Any expression of CD56 was considered for tumor exclusion from the present study, as referred.

Ki-67 LI scoring

A binomial cut-off for ki-67 LI was defined at 30%, in accordance with previous studies, reporting this value as cut-off for prognosis assessment in pulmonary carcinomas instead of median ki-67 LI value, which is not clinically relevant according to literature [18].

PD-L1 scoring

Immunohistochemical PD-L1expression was scored after PD-L1 staining stratification through negative (0% expression in TCs), + (1-5%), ++ (5-50%) and +++ (>50%). To make pathologists work reproducible, this estimation used the aforementioned

four-point cut-off in order to approach the thresholds routinely employed in diagnostic settings, without interfering with criteria for pembrolizumab prescription [19,20].

The binary PD-L1 expression score currently indicated for immunotherapy with pembrolizumab in advanced/metastatic lung cancer, establishing tumors with PD-L1 score of ≥ 1% but less than 50% to follow second-line therapy after one prior chemotherapy regimen and first-line treatment when 50% or more positive TCs are recognized in biopsies, had been clearly sustained in the reports [5,9].

In this study, cases with PD-L1 positive TCs (>1%) were then separated by +, ++ and +++ scores, and tumors with negative score (no stained TCs) formed another group.

The interpretation of PD-L1 immunostaining, as well as IHC correlations and final tumor diagnosis based in both histopathology predominant pattern and IHC panel expression, are represented in Table 2 and illustrated in Figure 1. Carcinoma NOS diagnosis was consistent with representative bronchial biopsy cases where TTF1 and CK5/6 had no expression in TCs expressing CK7, with or without vimentin expression and without defined pattern, where giant and fusiform cells were also absent (Table 1).

	SQC (n=16)	ADC (n=64)	ADCSQC (n=7)	Carcinoma NOS (n=3)	Pleomorphic (n=7)	All patients (n=97)
				Age		
≤ 68	7	30	4	3	5	49
> 68	9	34	3	0	2	48
				Gender		
Male	14	47	6	3	5	75
Female	2	17	1	0	2	22
			В	iopsy type		
Bronchial	12	25	4	3	1	45
Transtho- racic	3	32	1	0	3	39
Surgical	1	5	2	0	3	11
Pleural	0	2	0	0	0	2

Table 1: Patients and sampling characteristics distribution according to lung carcinomas histopathological classification. SQC squamous cell carcinoma, ADC adenocarcinoma, ADSQC adenosquamous carcinoma, NOS not otherwise specified

Tumoral stroma classification

Tumoral stroma subdivision was performed into four groups by light microscopy, following observation of bronchopulmonary carcinomas, in accordance with criteria adopted in previous studies [21-23].

Tumoral stroma was then classified as lymphocytic-predominant/immune-inflamed (when the infiltration of tumor-associated lymphocytes was predominant in the tumor stroma, positioned in the proximity to TCs), fusiform cells predominance, mixed type (when a balance between lymphocytes and fusiform cells was present) and lepidic. The lepidic pattern was represented in a transthoracic biopsy of one mucinous ADC, where TCs proliferated along the surface of intact or enlarged alveolar walls, consistent with lepidic tumoral pattern defined in WHO 2015/2020 criteria for ADCs. Stromal classification of ADC and SQC samples is described in Table 2.

Statistical analysis

Statistical analysis was performed using SPSS statistics 26.0 software for Windows (SPSS, Chicago, USA). Descriptive statistics included median with range for continuous variables, and count and frequency for categorical variables. Associations between PD-L1 expression and stratified PD-L1 score with clinicopathological variables, IHC markers and stromal subtype followed a multistep statistical approach. Firstly, the existence of association between the binary PD-L1 expression and these variables was analyzed using Pearson's χ 2 test and Fisher's exact test. Secondly, these tests were applied in order to investigate the association between the stratified PD-L1 staining (negative, +, ++ or +++) and the parameters that were significantly associated with binary PD-L1 expression. Finally, a logistics regression was performed to ascertain the effects of PD-L1 staining stratification on the likelihood of positivity of IHC

markers selected in the previous tests. P<0.05 were considered statistically significant.

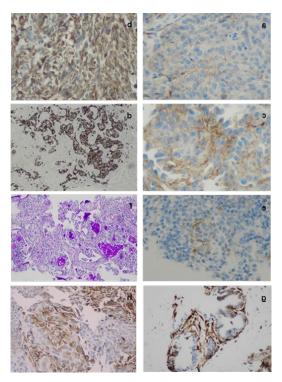


Figure 1: PD-L1 22C3 Dako immune expression is scored in routine Pathology following tumor cells complete and/or incomplete cytoplasmatic membrane immunostaining independent from intensity-squamous cell carcinoma with malignant spindle cells suggesting pleomorphic carcinoma in bronchial biopsy was scored with PD-L1 of 60%, x200 (a), sustained by CK5/6 expression, x400 (b); adenocarcinoma with relevant solid pattern in transthoracic biopsy with PD-L1 ≥ 50%, x400 (c) and cytoplasmatic CK7 expression, x100 (d); transthoracic biopsy of mucinous adenocarcinoma with PD-L1 5% score, x400 (e) and PAS-D mucin demonstration, x200 (f); transthoracic biopsy of adenocarcinoma-acinar pattern (CK7/TTF1 3+) with >50% vimentin expression in tumor cells, x200 (g) and vimentin >50% expression in bronchial biopsy of keratinizing squamous cell carcinoma, x400 (h).

RESULTS

PD-L1 expression and male gender patient tumors

PD-L1 positivity reported in **Table 2** concerned 56 cases, where 17 cases were classified as <5% expression in TCs, 10 cases between 5-50% and 29 cases over 50%, with complete/incomplete cytoplasmatic membrane staining.

PD-L1 positive expression was significantly associated with male gender (p=0.028): 48 of the 56 samples positive for PD-L1

expression were found among male individuals, while among the 22 female patient samples, 14 tumors were scored as PD-L1-negative. However, gender was not significantly associated with the stratified PD-L1 score (Table 3).

ADC solid pattern and higher PD-L1 expression

Among ADC cases evaluated for PD-L1 positivity, 21 of the 23 cases with solid pattern expressed PD-L1; 11 of the 12 acinar ADC cases and 13 of the 22 mucinous ADC cases were negative for PD-L1 expression (Supplementary Table 2).

Table 2: Immunohistochemical and stromal characterization of SQC and ADC biopsies. SQC squamous cell carcinoma, ADC adenocarcinoma, KTN keratinizing, non-KTN non keratinizing, LI labeling index

		SQC			ADC	
	KTN	non-KTN	Solid	Micropapillary	Acinar	Mucinous
	(n=7)	(n=9)	(n=23)	(n=7)	(n=12)	(n=22)
			CK7			
Positive	0	3	23	7	12	22
Negative	7	6	0	0	0	0
			TTF1			
Positive	0	0	22	7	10	17
Negative	7	9	1	0	2	5
			PAS-D			
Positive	0	0	0	0	1	22
Negative	7	9	23	7	11	0
			CK5/6			
Positive	7	9	1	0	0	0
Negative	0	0	22	7	12	22
			Vimentin			
Positive	2	2	6	6	2	5

Negative	5	7	17	1	10	17	
			CD56				
Positive	0	0	0	0	0	0	
Negative	7	9	23	7	12	22	
			Ki-67 LI				
≤30%	2	0	2	2	2	6	
>30%	5	9	21	5	10	16	
			PD-L1 expression				
> 50%	2	3	10	5	1	3	
5 - 50%	0	2	6	0	0	1	
< 5%	1	1	5	0	0	5	
Negative	4	3	2	2	11	13	
			Stroma subtype				
Immune-inflamed	1	1	5	0	1	1	
Mixed	2	2	10	5	4	8	
Fusiform	4	6	8	2	7	12	
Lepidic	0	0	0	0	0	1	

Table 3: Patients, pathological factors and PD-L1 positivity in tumor cells. Data presented as n(%). Pearson's $\chi 2$ and Fisher's exact test results. IHC immunohistochemistry, LI labeling index, pos. positive

	Namativa	PD-L1 pos. cases		Stratification of PD-L1 pos. cases			
	Negative	Total	Decelor	+	++	+++	D l
	(n=41)	(n=56)	P value	(n=17)	(n=10)	(n=29)	P value
Gender			0.028				0.131
Male	27 (65.85)	48 (85.71)		15 (88.24)	9 (90.00)	24 (82.76)	
Female	14 (34.15)	8 (14.29)		2 (11.76)	1 (10.00)	5 (17.24)	
IHC markers							
Vimentin			0.018				0.049
Positive	8 (19.51)	24 (42.86)		5 (29.41)	5 (50.00)	14 (48.28)	at
Negative	33 (80.49)	32 (57.14)		12 (70.59)	5 (50.00)	15 (51.72)	at
Ki-67 LI			0.029				0.026
≤ 30%	11 (26.83)	5 (9.26)		4 (25.00)	0 (0.00)	1 (3.57)	
>30%	30 (73.17)	49 (90.74)		12 (75.00)	10 (100.00)	27 (96.43)	
Stroma sub- type			0.151				0.506
Immune-in- flamed	1 (2.50)	9 (16.07)	at	3 (17.65)	1 (10.00)	5 (17.24)	
Mixed	14 (35.00)	21 (37.50)	at	5 (29.41)	4 (40.00)	12 (41.38)	
Fusiform	24 (60.00)	25 (44.64)	at	8 (47.06)	5 (50.00)	12 (41.38)	
Lepidic	1 (2.50)	1 (1.79)	at	1 (5.88)	0 (0.00)	0 (0.00)	

SQC variable PD-L1 expression

In the 16 SQC samples, 3 cases expressed CK7 and of the 13 SQC CK7-negative samples, 8 expressed PD-L1, and 4 of these cases had PD-L1 expression over 50% in TCs (Supplementary Table 3).

Vimentin expression as an independent marker for immunotherapy selection

Relationship between vimentin expression and PD-L1 positive expression was also significant (p=0.018) (Table 3). Vimentin expression was positive in 32 cases, 24 of which also showed positive PD-L1 expression; and among the 41 PD-L1-negative samples, 33 cases were also negative for vimentin expression. The stratified PD-L1 score was found significantly associated with vimentin expression (p=0.049): in 24 vimentin-positive/PD-L1-positive samples, 14 had PD-L1 expression over 50% in TCs (Table 3). Vimentin was also significantly associated with

histopathological subtyping (p=0.037): 5 of the 7 pleomorphic carcinomas had vimentin expression over 50% in TCs (Supplementary Table 4).

Table 4: Risk analysis of vimentin positivity and ki-67 LI>30%. Logistics regression results, LI labeling index, Ref. reference, OR odds ratio, CI confidence interval.

		PD-L1 stratified score				
	Negative	+	++	+++		
Vimentin ex	pression					
OR	Ref.	1.72	4.13	3.85		
95% CI	-	0.47-6.29	0.957-17.77	1.33 - 11.13		
P value	-	0.413	0.057	0.013		
Ki-67 LI > 30%						
OR	Ref.	1.1	592340776	9.9		

95% CI - 0.29-4.14 - 1.20 - 81.83 P value - 0.888 - 0.033

A logistic regression was performed to ascertain the effects of PD-L1 expression on the likelihood that samples were positive for vimentin expression. Samples with more than 50% of PD-L1 stained TCs were 3.85 times more likely to be vimentin-positive than PD-L1-negative specimens (OR=3.85; p=0.013) (Table 4).

Ki-67 30% cut-off applicable in ADCs

A significant association was found between ki-67 LI and PD-L1 expression (p=0.029), where 49 of 54 positive PD-L1 cases had ki-67 LI>30% (Table 3).

The stratified PD-L1 score was also significantly associated with ki-67 LI (p=0.026), as 37 from 38 samples with PD-L1 score >5% presented ki-67 LI>30% (Table 3).

A logistic regression was used to determine the relationship between PD-L1 expression and ki-67 LI>30%. Cases with PD-L1 expression over 50% in TCs showed a 9.90 times higher probability of having ki-67 LI>30%, versus PD-L1 negative tumors (OR=9.90; p=0.033) (Table 4).

Immune-inflamed stroma and PD-L1 expression correlation Patients' age, immunohistochemistry panel, PAS-D and carcinoma histopathological subtyping did not show a significant association with PD-L1 expression (Supplementary Table 5).

A tendency to PD-L1 positive expression came up in lymphocytic-predominant/immune-inflamed stroma samples (p=0.151), where 9 of 10 samples with this stroma subtype showed positive PD-L1 expression in TCs (Table 3).

DISCUSSION

As heterogenous diseases, either at cellular and histopathological perspective, with distinct diagnostic, prognostic and therapeutic features [24], ADC and SQC keep being the most prevalent bronchopulmonary carcinomas, responsible for approximately 50% and 30% of cases, respectively [3].

With 5-year survival rate still approximately 15% [3], near 30% of patients with tumors in non-surgical stages have mutations amenable to targeted therapy [25], and PD-1/PD-L1 inhibitors have been prolonging patients survival with a more acceptable toxicity profile, proving undoubted superiority over chemotherapy and targeted therapy in terms of efficacy [10,26].

Durable host immune anti-neoplasm responses and long-term remissions of several tumor types proved favorable benefit-to-risk of anti-PD therapy [9,27]. For advanced carcinomas without EGFR/ALK mutations, European Medicines Agency (EMA) and FDA approved pembrolizumab monotherapy after PD-L1 score ≥ 50% or in combination with pemetrexed and platinum chemotherapy in carcinomas other than SQCs as first-line treatment, and as monotherapy for ADCs with PD-L1 expression between 1% and 50% after at least one prior chemotherapy regimen. [5,9,25].

Relationship between PD-L1 expression and gender remains contradictory [28,29]. Concerning a limited series, our findings demonstrated that PD-L1 expression was significantly associated with gender, as 48 of 56 positive PD-L1 cases belonged to male patients, which might be due to unrecognized hormonal differences between both genders and their influence on PD-L1 expression, deserving further research on the matter as there is currently no solid explanation for this finding.

It is becoming evident that histopathological subtyping relates to PD-L1 expression in TCs, particularly among solid ADC subgroup, still with worse prognosis. Driver et al. and Mandarano et al. demonstrated that lung ADCs defined by PD-L1 expression in TCs and tumor-infiltrating immune cells correlated with solid pattern, while acinar, mucinous and papillary patterns presented lower PD-L1 expression in TCs [4,30]. As in literature [24,28], our study confirmed that positive PD-L1 expression was relevant in solid pattern of ADCs (21/23), defining a clear and expected relationship between PD-L1 expression and an aggressive histopathological subtype, based on PD axis properties of promoting tumor survival and immune evasion.

EGFR mutations and ALK rearrangements keep being rare in advanced lung SQC, while immunotherapeutic strategies have been particularly effective [31,32]. In CheckMate 017 trial, nivolumab improved survival, PFS and response rate versus docetaxel [33]. Following progression after first-line chemotherapy, PD-L1 inhibitors are the preferred treatment, according to U.S. National Comprehensive Cancer Network (NCCN) guidelines [31].

Cytokeratin 7, glandular/anterior gut differentiation marker present in normal glandular and transitional epithelium but not in squamous epithelium and expressed in 60-100% of ADCs [34,35], is used to subclassify lung SQC into two groups: pure SQC (CK7-negative), and non-pure SQC with CK7 expression, according to WHO 2015/2020 criteria for lung tumors. In considered pure SQC subgroup, EGFR and ALK mutations are almost absent and targeted therapy is much more limited, while for non-pure SQCs, molecular pathology may define therapy [36].

In our results, 8 cases out of 13 CK7-negative SQCs expressed PD-L1, with 4 cases over 50% of positive TCs. To our knowledge, this study is the first to document the tendency to high PD-L1 expression in pure lung SQC cases (CK7-negative), which needs to be further characterized in future with a greater number of sample biopsies for a more personalized application of PD-1/PD-L1 immune checkpoint inhibitors in so-called pure SQCs.

The applicability of tumor microenvironment (TME) as a diagnostic, prognostic or predictive biomarker in bronchopulmonary carcinomas seems to correlate with tumorigenesis, heterogeneity, resistance to immunotherapy and tumor progression [2,37], and also stromal cells may express the ligand PD-L1, with still unclear meaning [37].

The tendency of high PD-L1 expression in TCs among cases with lymphocytic-predominant/immune-inflamed stroma was 9/10 samples. Our results might be explained as the induction of TCs PD-L1 expression is mediated by interferon- γ produced by T lymphocytes present in the TME [38]. Tumor-infiltrating lymphocytes (TILs) have also been proposed as a biomarker of response for PD-1/PD-L1 inhibition therapy [39,40]. Anti-PD therapy seems also less effective in non-inflamed tumors (low lymphocyte infiltration/PD-L1 expression) with increased levels of transforming growth factor- β (TGF- β), inducer of resistance to anti-PD-L1 therapy, and fusiform cells rich stroma [41].

Epithelial-mesenchymal transition (EMT) with epithelial cells becoming mesenchymal cells by losing cell-cell adhesion and polarity and acquiring invasive/migratory properties, might contribute to drug resistance and hence poor prognosis [2,37,42,43]. E-cadherin downregulation and overexpression of vimentin, N-cadherin, α -actin and fascin depending on EMT

genes, such as snail family transcriptional repressor 2 (Slug), twist family bHLH transcription factor 1 (TWIST), zinc finger E-box binding homeobox 1 (ZEB1), nuclear-translocated β -Catenin and TGF- β , have been associated with TKI resistance, namely to EGFR-TKIs [29,40,44].

To Kim et al., PD-L1 expression may be responsible for EMT oncogenesis and immune evasion during tumor development [45], contradictory with EMT-induced PD-L1 expression in pulmonary carcinomas [46]. PD-L1 and EMT bidirectional crosstalk has since then been proposed to promote tumor aggressiveness [45,47]. Neurotrophic tyrosine receptor kinase (NTRK) gene rearrangements present in 0.1% to 1% of lung carcinomas, assessed by NGS and with effective targeted therapy [48,49], are associated with microscopically high grade features and less differentiated phenotype in mesenchymal tumors [49], needing further studies to be correlated with EMT phenotype in carcinomas, where vimentin and other EMT markers expression might become relevant.

A significant association was found between PD-L1 expression and high vimentin expression in TCs, with 24 of 32 vimentin-positive cases expressing PD-L1, versus 33 of 41 PD-L1-negative samples without vimentin expression. This result was consistent with previous observations that PD-L1 expression was positively correlated with vimentin expression and EMT phenotype in lung ADC, extrahepatic cholangiocarcinoma, breast carcinoma and head/neck/esophageal squamous carcinoma, suggesting that tumors with EMT status stand as potential targets for immunotherapy agents [45,47]. In fact, Ancel et al. proposed vimentin as canonical marker and actor of EMT [29], verified in this series in 24 vimentin-positive/PD-L1-positive cases where 14 had PD-L1 expression in over 50% TCs, becoming a frequent event upon increasingly higher PD-L1 expression, significantly leading the risk of 3.85 times higher expression among cases with more than 50% PD-L1 stained TCs, versus PD-L1 negative samples.

Proliferation marker ki-67 keeps being associated with tumor aggressiveness and metastization in solid tumors [50]. A significant association was present between ki-67 LI and both positive PD-L1 expression and stratified PD-L1 score, as 49 of 54 PD-L1-positive cases had ki-67 LI>30% and 37 from 38 samples with more than 5% of PD-L1 stained TCs showed ki-67 LI>30%. Association between PD-L1 status and tumor cell proliferation was also confirmed by the tendency of PD-L1 positivity in the solid pattern of ADC samples, in accordance with literature [19,51], still contradictory regarding association between PD-L1 expression and ki-67 LI in SQCs [51]. Similarly to vimentin, the risk of ki-67 LI>30% is 9.90 times higher in samples with more than 50% of PD-L1 stained TCs, versus PD-L1 negative specimens.

To the best of our knowledge, our study was the first to investigate the relationship between PD-L1 expression and EMT status, evaluated by vimentin included in IHC panel applied in Pathology practice, with perspective of risk analysis. Similarly to previous investigations [43,45], EMT status by vimentin staining can be relevant in selecting patients more likely to have higher TCs PD-L1 expression. Favorable response to PD-1/PD-L1 immune checkpoint blockade in bronchopulmonary carcinomas would be then more accurate based on two biomarkers expressed in TCs, namely in follow-up of targeted therapy acquired resistance [42]. Also, as 10-20% of unselected

patients with advanced carcinomas benefit from anti-PD therapy [11,27,39,41], combined therapy of PD-1/PD-L1 inhibitors with EMT targeted therapies might become an ultimate therapeutical option and vimentin might work as a fundamental biomarker.

Combination of mitogen-activated protein kinase kinase (MEK) inhibitors with PD-L1 inhibitors improved tumor regression, where MEK inhibitors may sensitize TCs to immunotherapy agents [47]. Mechanistic target of rapamycin (mTOR) promotes EMT phenotype and immune evasion through upregulation of PD-L1 expression and the effect of mTOR inhibition combined with PD-L1 blockade was also reported in preclinical lung cancer trials [47]. Finally, combination of PD-1 inhibitors with EGFR TKIs in PD-L1-positive carcinomas with EGFR activating mutations raised promising results in preclinical trials, as EGFR activation up-regulated PD-L1 expression, probably making these tumors more susceptible to PD-1/PD-L1 blockade therapy [25,42,47]

Limitations of this study, concerning 97 biopsies, and the utility of the applied IHC panel, deserve replication with additional EMT markers, to support EMT-phenotype as a new potential predictive biomarker to define immunotherapy in pulmonary carcinomas. Research on combined therapies with EMT targeted agents and PD-L1 inhibitors to improve patient's outcome and survival deserve to be implemented.

CONCLUSION

An exercise PD-L1 expression was significantly associated with vimentin expression and ki-67 LI>30% and this association was maintained when stratified according to increasing intervals of PD-L1 expression score.

PD-L1 positive samples with more than 50% stained TCs had a significantly increased risk of expressing vimentin and presenting with high proliferation status defined by ki-67 Ll>30%.

Consequently, ki-67 LI>30% and vimentin expression may become rationale biomarkers that can be used to identify tumors more likely to benefit from PD-1/PD-L1 axis blockade, overcoming the limitations of single PD-L1 IHC scoring due to tumoral heterogeneity and high staged carcinomas associated with resistance to targeted therapy.

Vimentin expression and ki-67 LI may also complement the evaluation of TMB as cost-effective and available markers. Reinforcing EMT targeted therapy agents combined with PD-L1 inhibitors in bronchopulmonary carcinomas, an EMT phenotype is raising as a promising field for future research.

ACKNOWLEDGEMENTS

The authors thank Dr. Ana Ladeirinha (Institute of Anatomical and Molecular Pathology, Faculty of Medicine, University of Coimbra, Portugal) for support with statistical analysis and execution of immunohistochemistry.

CONFLICT OF INTEREST

The authors report no conflicts of interest.

FUNDING

The authors did not receive funding for conducting this study

DATA AVAILABILITY

Data sets from this study are available upon request from the corresponding author.

CODE AVAILABILITY

Not applicable.

ETHICS APPROVAL

The study fulfilled the rules for archival retrospective study defined by the Faculty of Medicine of the University of Coimbra Ethical Committee.

CONSENT TO PARTICIPATE

Not applicable.

CONSENT FOR PUBLICATION

Not applicable.

REFERENCES

- Lin A, Wei T, Meng H, Luo P, Zhang J (2019) Role of the dynamic tumor microenvironment in controversies regarding immune checkpoint inhibitors for the treatment of nonsmall cell lung cancer (NSCLC) with EGFR mutations. Mol Cancer 18(1):139.
- 2. Altorki NK, Markowitz GJ, Gao D (2019) The lung microenvironment: An important regulator of tumour growth and metastasis. Nat Rev Cancer 19(1):9-31.
- Osmani AL, Askin F, Gabrielson E, Li QK (2017) Recent large scale genomic studies from the Clinical Lung Cancer Genome Project have SC. Semin Cancer Biol.
- Mandarano M, Bellezza G, Belladonna ML (2019) Assessment of TILs, IDO-1, and PD-L1 in resected non-small cell lung cancer: An immunohistochemical study with clinicopathological and prognostic implications. Virchows Arch 474(2):159-168.
- Du Rusquec P, De Calbiac O, Robert M, Campone M, Frenel JS (2019) Clinical utility of pembrolizumab in the management of advanced solid tumors: An evidence-based review on the emerging new data. Cancer Manag Res 11:4297-4312.
- Meléndez B, Van Campenhout C, Rorive S, Remmelink M, Salmon I, et al. (2018) Methods of measurement for tumor mutational burden in tumor tissue. Transl Lung Cancer Res 7(6):661-667.
- Hirsch FR, McElhinny A, Stanforth D (2017) PD-L1 immunohistochemistry assays for lung cancer: Results from phase 1 of the blueprint PD-L1 IHC assay comparison project. J Thorac Oncol 12(2):208-222.
- Tsao MS, Kerr KM, Kockx M (2018) PD-L1 Immunohistochemistry Comparability Study in Real-Life Clinical Samples: Results of Blueprint Phase 2 Project. J Thorac Oncol 13(9):1302-1311.
- Munari E, Zamboni G, Lunardi G (2018) PD-L1 expression heterogeneity in non–small cell lung cancer: Defining criteria for harmonization between biopsy specimens and

- whole sections. J Thorac Oncol 13(8):1113-1120.
- Teixidó C, Vilariño N, Reyes R, Reguart N (2018) PD-L1 expression testing in non-small cell lung cancer. Ther Adv Med Oncol 10:1-17.
- 11. Kerr KM (2018) The PD-L1 immunohistochemistry biomarker: Two steps forward, one step back? J Thorac Oncol 13(3):291-294.
- Chen Y, Liu Q, Chen Z (2019) PD-L1 expression and tumor mutational burden status for prediction of response to chemotherapy and targeted therapy in non-small cell lung cancer. J Exp Clin Cancer Res 38(1):1-14.
- 13. Chan TA, Yarchoan M, Jaffee E. (2019) Development of tumor mutation burden as an immunotherapy biomarker: Utility for the oncology clinic. Ann Oncol 30(1):44-56.
- 14. Kazdal D, Endris V, Allgäuer M (2019) Spatial and temporal heterogeneity of panel-based tumor mutational burden in pulmonary adenocarcinoma: separating biology from technical artifacts. J Thorac Oncol 14(11):1935-1947.
- 15. Rizvi NA, Hellmann MD, Snyder A (2015) Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science (80-) 348(6230):124-128.
- 16. Zheng M (2016) Classification and Pathology of Lung Cancer. Surg Oncol Clin N Am 25(3):447-468.
- Brown NA, Aisner DL, Oxnard GR (2018) Precision medicine in non–small cell lung cancer: current standards in pathology and biomarker interpretation. Am Soc Clin Oncol Educ B (38):708-715.
- 18. Jakobsen JN, Sørensen JB (2013) Clinical impact of ki-67 labeling index in non-small cell lung cancer. Lung Cancer 79(1):1-7.
- 19. Pawelczyk K, Piotrowska A, Ciesielska U (2019) Role of PD-L1 expression in non-small cell lung cancer and their prognostic significance according to clinicopathological factors and diagnostic markers. Int J Mol Sci 20(4):1-15.
- Rimm DL, Han G, Taube JM (2017) A prospective, multi-institutional, pathologist-based assessment of 4 immunohistochemistry assays for PD-L1 expression in non–small cell lung cancer. JAMA Oncol 3(8):1051-1058.
- 21. Silva MR, Alarcão A, Ferreira T (2016) Evaluation of HER2 by automated FISH and IHC in gastric carcinoma biopsies. Int J Biol Markers 31(1):e38-e43.
- 22. Iwata H (2016) Adenocarcinoma containing lepidic growth. J Thorac Dis 8(9):E1050-E1052.
- 23. Chen DS, Mellman I (2017) Elements of cancer immunity and the cancer-immune set point. Nature 541(7637):321-330.
- 24. De Sousa VML, Carvalho L (2018) Heterogeneity in Lung Cancer. Pathobiology 85(1-2):96-107.
- 25. Yuan M, Huang LL, Chen JH, Wu J, Xu Q (2019) The emerging treatment landscape of targeted therapy in non-small-cell lung cancer. Signal Transduct Target Ther 4(1).
- 26. Blumenthal GM, Bunn PA, Chaft JE (2018) Current status and future perspectives on neoadjuvant therapy in lung cancer. J Thorac Oncol 13(12):1818-1831.
- 27. Zou W, Wolchok JD, Chen L (2016) PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: Mechanisms,

- response biomarkers, and combinations. Sci Transl Med 8(328).
- 28. Miyazawa T, Marushima H, Saji H (2019) PD-L1 expression in non-small-cell lung cancer including various adenocarcinoma subtypes. Ann Thorac Cardiovasc Surg 25(1):1-9.
- 29. Ancel J, Birembaut P, Dewolf M (2019) Programmed death—ligand 1 and vimentin: A tandem marker as prognostic factor in NSCLC. Cancers (Basel) 11(10):1-14.
- Driver BR, Miller RA, Miller T (2017) Programmed death ligand-1 (pd-l1) expression in either tumor cells or tumor-infiltrating immune cells correlates with solid and high-grade lung adenocarcinomas. Arch Pathol Lab Med 141(11):1529-1532.
- 31. Socinski MA, Obasaju C, Gandara D (2018) Current and emergent therapy options for advanced squamous cell lung cancer. J Thorac Oncol 13(2):165-183.
- 32. Jett JR, Carr LL (2013) Targeted therapy for non-small cell lung cancer. Am J Respir Crit Care Med 188(8):907-912.
- 33. Hirsch FR, Scagliotti G V, Mulshine JL (2017) Lung cancer: Current therapies and new targeted treatments. Lancet 389(10066):299-311.
- Wang J, Wang X, Xu X, Li S, Xiu-lan (2020) NapsinA, CD56, Syn and CgA in biopsy Specimen of squamous cell carcinoma, adenocarcinoma and small cell lung carcinoma. Int J Morphol 38(2):247-251.
- 35. Jafarian AH, Gharib M, Mohammadian Roshan N, Sherafatnia S, Omidi AA, et al. (2017) The diagnostic value of TTF-1, P63, HMWK, CK7, and CD56 immunostaining in the classification of lung carcinoma. Iran J Pathol 12(3):195-201.
- 36. Huang Y, Wang R, Pan Y (2016) Clinical and genetic features of lung squamous cell cancer in never-smokers. Oncotarget 7(24):35979-35988.
- 37. Valkenburg KC, De Groot AE, Pienta KJ (2018) Targeting the tumour stroma to improve cancer therapy. Nat Rev Clin Oncol 15(6):366-381.
- 38. Seliger B (2019) Basis of PD1 / PD-L1 Therapies. J Clin Med 8:1-14.
- 39. Li HY, McSharry M, Bullock B (2017) The tumor microenvironment regulates sensitivity of murine lung tumors to PD-1/PD-L1 antibody blockade. Cancer Immunol Res 5(9):767-777.

- 40. Zacharias M, Brcic L, Eidenhammer S, Popper H (2018) Bulk tumour cell migration in lung carcinomas might be more common than epithelial-mesenchymal transition and be differently regulated. BMC Cancer 18(1):1-13.
- 41. Zhao S, Ren S, Jiang T (2019) Low-dose apatinib optimizes tumor microenvironment and potentiates antitumor effect of PD-1/PD-L1 blockade in lung cancer. Cancer Immunol Res 7(4):630-643.
- 42. Denisenko TV, Budkevich IN, Zhivotovsky B (2018) Cell death-based treatment of lung adenocarcinoma article. Cell Death Dis 9(2).
- 43. Jung AR, Jung CH, Noh JK, Lee YC, Eun YG (2020) Epithe-lial-mesenchymal transition gene signature is associated with prognosis and tumor microenvironment in head and neck squamous cell carcinoma. Sci Rep 10(1):1-11.
- 44. Zhu X, Chen L, Liu L, Niu X (2019) EMT-Mediated Acquired EGFR-TKI Resistance in NSCLC: Mechanisms and Strategies. Front Oncol 9(October):1-15.
- 45. Kim S, Koh J, Kim MY (2016) PD-L1 expression is associated with epithelial-to-mesenchymal transition in adenocarcinoma of the lung. Hum Pathol 58:7-14.
- 46. Funaki S, Shintani Y, Ura TK, Kanzaki R, Minami M, et al. (2017) Chemotherapy enhances programmed cell death 1/ligand 1 expression via TGF-β induced epithelial mesenchymal transition in non-small cell lung cancer. Oncol Rep 38(4):2277-2284.
- 47. Jiang Y, Zhan H (2020) Communication between EMT and PD-L1 signaling: New insights into tumor immune evasion. Cancer Lett 468:72-81.
- 48. Farago AF, Taylor MS, Doebele RC (2018) Clinicopathologic features of non–small-cell lung cancer harboring an NTRK gene fusion. JCO Precis Oncol (2):1-12.
- 49. Antonescu CR, Dickson BC, Swanson D (2019) Spindle cell tumors with RET gene fusions exhibit a morphologic spectrum akin to tumors with NTRK gene fusions. Am J Surg Pathol 43(10):1384-1391.
- 50. Mitin T, Choudhury A (2019) The role of biomarkers in bladder preservation management of muscle-invasive bladder cancer. World J Urol 37(9):1767-1772.
- 51. Shimoji M, Shimizu S, Sato K (2016) Clinical and pathologic features of lung cancer expressing programmed cell death ligand 1 (PD-L1). Lung Cancer 98:69-75.