ROUND TABLE

Tumor Destruction and *In Situ* Delivery of Antigen Presenting Cells Promote Anti-Neoplastic Immune Responses: Implications for the Immunotherapy of Pancreatic Cancer

Patrizia Rovere-Querini, Angelo A Manfredi

Cancer Immunotherapy and Gene Therapy Program, Clinical Immunology Unit, H San Raffaele Scientific Institute and Vita-Salute San Raffaele University. Milan, Italy

Summary

Antigen presenting cells (APCs) activate helper and cytotoxic T cells specific for antigens expressed by tissue cells, including neoplastic cells. This event occurs after the antigen transfer from tissue cells to APC, and is referred to as "cross-presentation". The number and the state of activation of APC in the tumor control the outcome of crosspresentation, including the establishment of protective immune responses. Cell death favors cross-presentation. Cancer cells normally die, either spontaneously or as a consequence of targeted therapies. The transfer of tumor antigens from dying tumor cells to APCs in vivo, exploiting the crosspresentation pathway, has the potential of yielding novel immunotherapeutic strategies. Their success will depend on at least two factors: the induction of synchronized cell death in the tumor, and the recruitment of activated dendritic cells in the tumor. Under normal conditions pancreatic cancer represents a privileged environment; its profound chemoresistance reflects limited apoptosis after chemotherapy. Moreover, it usually contains only a few cells endowed APC with function. Endoscopic ultrasonography offers attractive possibilities of circumventing this privilege, including the delivery of ultrasound, radiofrequency or radiation in order to destroy the tumor and the

delivery *in situ* of autologous APC or appropriate chemotactic signals. In general, loco-regional approaches offer the possibility of using the tumor of each patient as a complex antigen source, thus limiting the risk of tumor escape and reducing the need for extensive *ex vivo* handling of the neoplasm and of the patient APCs.

Tumor Immunogenicity and Antigen Presenting Cells

The diagnosis of pancreatic cancer is often established in patients who have advanced diseases, with approximately half of them having metastases and the majority of the rest having locally diffused, unresectable disease. Moreover, this tumor is highly resistant to quite chemotherapy [1], with an unsatisfactory systemic mode of treatment and an extremely low survival rate. Novel therapies are sorely needed [1, 2, 3]. In recent vears we have developed a better understanding of the interaction between the tumor and the host. Neoplastic cells express specific antigens which can be recognized by immune cells, leading to regression of the neoplasm and patient cure [4, 5]. Immunemediated regression implies that tumorspecific T cells have been properly activated, a task accomplished in vivo by specialized immune cells, referred to as antigen presenting cells (APCs).

Dendritic cells (DCs) are the most potent APCs. They originate from CD34+ bone marrow progenitors and reach peripheral organs via the blood. At the periphery, they develop to immature DCs, specialized in sampling tissue antigens via macropinocytosis, phagocytosis or receptormediated endocytosis. Primary proinflammatory signals switch the main function of DCs from antigen-uptake to presentation. Maturing DCs rearrange the actin-based cytoskeleton, acquire the sensitivity of attracting signals generated in the lymph nodes, where they migrate via afferent lymphatics. DCs complete the processing of the antigenic material on the way to the lymph node and up-regulate the expression of the molecules involved in the antigen presentation to the T cells [6, 7].

DCs infiltrate several tumors, including melanomas and carcinomas of the lung, stomach, prostate thyroid and breast. Tumor cells, by release of factors such as IL-6, IL-10, and vascular endothelial growth factor, influence DCs, inhibiting their maturation and their ability to migrate to secondary lymphoid organs and activate T cells [8]. Tumor associated DCs are indeed often endowed а low allostimulatory capacity. with particularly if isolated from the progressing metastatic lesions. Moreover, at least in breast carcinomas, DCs are compartmentalized, with relatively large numbers of immature cells "trapped" in the lesion while mature DCs are preferentially found within the peritumoral areas [9]. A better survival rate after lymphadenectomy with a favorable clinical outcome and even the spontaneous regression of metastatic melanomas are associated with the infiltration of the tumor parenchyma by mature DCs, establishing close interactions in situ with tumor specific cytotoxic T cells [10].

Dendritic Cells, Cross-Presentation and Tumor Cell Death

The transfer of antigens expressed in neoplastic tissues to DCs for the productive

activation of T cells is a key event for the initiation of immune responses against the tumor. The transfer of intracellular antigens among living cells is a rare event. In contrast, cell death associates with the loss of the compartmentalization of intracellular components which are released in the microenvironment, where thev elicit inflammatory responses [11, 12]. Moreover, antigen presenting phagocytes efficiently internalize apoptotic tumor cells, process them in dedicated intracellular vesicles and cross-present their antigens in vivo and in vitro [13, 14, 15, 16]. Tumor-associated DCs apparently equally proficient are at internalizing dying tumor cells, processing them and cross-priming tumor specific T lymphocytes [17].

Tumor cell death is important for generating a proper phagocytic meal for antigen presenting DCs by feeding them with tumor antigens [18]. Tissue damage per se activates immune responses [19, 20, 21] with acute but generally transient immune responses against intracellular antigens [22, 23, 24]. The preferential recognition of antigens contained in dying cells is facilitated by the release of soluble ("immune factors endogenous adjuvants") which amplify and sustain the function of APCs and the establishment of T cell dependent immune responses, in situ and at a distance [25, 26]. Therefore, the massive death of tumor cells, overwhelming the phagocytic ability of local scavenger cells, pro-immune environment. generates а Immunostimulatory factors include uric acid [27] and heat shock proteins, whose expression is often deregulated in neoplastic factors tissues [28]. Other intracellular released by necrotic cells with similar functions still remain to be identified.

Both antigen release and the delivery of immunostimulatory signals to APCs possibly contribute to the results described using radiofrequency ablation in an experimental mouse system [29]. Radiofrequency ablation is minimally invasive, widely used in different clinical settings and results in the generation of large burden of tumor cell debris [30]. Radiofrequency ablation alone induced а specific immune response, dependent of the cellular arm of the immune system, capable of protecting a fraction of mice against a second challenge with a lethal dose of living tumor cells [29]. This result suggests that even procedures commonly used in clinical practice which result in the synchronized death of tumor cells, have the potential of initiating a specific immune response. It is, of course, quite clear that, in the absence of interventions aimed at amplifying and sustaining the elicited response, the treatment per se is unlikely to show any clinical benefit at a distance. Even in the model described in the study by Den Brok et al., only were about the 20% of the treated animals actually protected [29].

Several factors contribute to limiting the efficacy of the response. They include the lack of activated APCs in the tumor in the correct time-frame for internalizing dying tumor cells and cross-presenting their antigens. Moreover, ablative treatments induce the *in vivo* release of pro-inflammatory factors and the production of acute phase proteins, such as the short pentraxin Creactive protein, which is produced in the liver under the control of IL-6 [31]. Pentraxins bind to dying cells and control immunogenicity, regulating their their interaction with antigen presenting cells and controlling their interaction with key factors of the innate immune response, like the complement system [32, 33, 34, 35]. It is tempting to speculate that elevated levels of acute phase pentraxins, which possibly physiologically prevent the induction of autoimmunity after massive cell death [34]. contribute quenching also to the immunogenicity of dying tumors and limiting protective immunity after ablation of human cancers.

Taken together, these findings imply that several interacting factors are involved in the initiation of antineoplastic immunity, including the availability of APCs at the tumor site, their activation state, the extent of cell death taking place in the neoplasm, the possible recruitment of immunosuppressive circuits at the site of the tumor cell death.

Pancreatic Cancer as a Privileged Tissue

Extensive tumor cell death takes place in human cancer, either spontaneously or as a consequence of targeted therapies. The case of pancreatic cancer is peculiar; its poor prognosis is indeed associated with resistance to cytotoxic drugs, a phenomenon known as chemoresistance [1]. Of importance, pancreatic adenocarcinomas comprise an important mesenchymal stromal component. Continuous tumor-stroma interactions have been proposed to lead to the generation of anti-apoptotic gaseous soluble and messengers [36]. Chemoresistance could therefore associate to apoptosis resistance, resulting in the lack of cell death induction at the tumor site. A second factor possibly restricting the immunogenic potential of pancreatic cancer is the lack of DCs as compared to other neoplasms [37]. Both lack of cell death and lack of interaction between tumor and host APCs might contribute to poor T cell priming and failure of tumor rejection [5]. The reasons for the selective paucity of DCs in pancreatic cancer (and not, for example, in laryngeal, colorectal, lymphoma, uterine, gastric, papillary thyroid, or lung cancers, in which the presence of tumorassociated DCs correlates with improved survival [37]) are poorly understood. Theoretically, DC apoptosis could be induced by the tumor [38]. Indeed, the exposure of DCs propagated *in vitro* to the anti-apoptotic gaseous messenger, nitric oxide, both protects them from apoptosis in vivo and enhances their function in the tumor [39]. However, some evidence suggests that a defective homing of DCs to the tumor, more than the active induction of cell suicide of infiltrating DCs, is involved. Schmidt et al. developed an experimental model of pancreatic cancer based on the implantation of ductal pancreatic adenocarcinoma cells in healthy nice; this model nicely reproduces several features of the human disease, including the lack of infiltrating DCs [40]. These authors therefore relied on ultrasound-guided orthotopic inoculation to inject DCs into the growing tumors. DC survival was apparently

unaffected by the tumor environment *in vivo* while, provided that they had been previously loaded with tumor antigens, DCs generated a potent antineoplastic immune response, quite superior to the response elicited by DC vaccination s.c. or i.v.. These results suggest that, once transferred into the pancreatic cancer, DCs not only survive but have a conserved antigen presenting function.

APCs and Loco-Regional Strategies

Direct injection of DCs into the tumor lesion has been proposed as a reliable approach in experimental systems. This procedure minimizes the need of in vitro loading with tumor antigens, circumvents the problem of efficient homing systemically delivered DCs to the tumor, and can be efficiently combined with other approaches, including chemotherapy and radiotherapy [41, 42, 43]. However, alternative approaches exist to enrich the tumor with activated APCs, which may prove valuable in the clinical setting. For example, Furumoto et al. [44] directed circulating DCs to experimental tumors inducing the expression of the CCL20/MIP-3alpha chemokine by gene transduction or by directly injecting the recombinant protein into the tumor. Depending on the model used, the simple attraction of DCs in situ was sufficient to elicit a specific anti-tumor response, dependent on CD4 and CD8 T cells. However, the activation state of DCs is a limiting factor (see above) and the receptor for CCL20/MIP-3alpha is down-regulated during DC maturation. In more stringent models. the simultaneous injection of unmethylated cytosine-guanine rich sequences and immunostimulatory oligonucleotide motifs to activate DCs was necessary in order to obtain clinically relevant protection [44]. Several other factors contribute, including the immunosuppression generated by the tumor or by conditioned infiltrating leukocytes, such as IL-10 [45, 46], which are known to inhibit DC maturation and function. Not surprisingly, in other studies the reversal of these inhibitory

influences, for example by an IL-10 blockade, was also necessary in order to establish a protective antineoplastic immune response [47].

Conclusions

Most of the findings described until now were obtained in experimental models. In the future, verifying whether immunotherapeutic strategies based on these insights will provide an advantage in clinical settings will be a major challenge. Pancreatic carcinoma is a natural candidate, given the unsatisfactory results of the therapeutic tools currently available. Moreover, the lack of infiltrating DC seems to quite clearly lay a possible roadmap for future study. The poor induction of cell death in the context of the tumor by systemic therapy could possibly be circumvented by ablative approaches, which result in efficient destruction of the targeted tissues (e.g. Chan et al. [48]). Endoscopic ultrasonography (EUS) is, at present, a powerful tool for imaging and substantiating diagnosis. However, its possibilities for antitumor therapy include the local destruction of tumor cells by ultrasound, radiofrequency or radiation [49]. The same approach may be exploited to locally deliver cells (including APC) or pharmacological agents (including drugs and chemokines). A creative use of the possibilities of EUS [49] will prove valuable for immunotherapy, increasing the possibility of a successful outcome of novel vaccination procedures.

Keywords Apoptosis; Dendritic Cells; Endosonography; Immunity, Natural; Necrosis

Abbreviations APC: antigen presenting cell; DC: dendritic cell

Acknowledgments The support of the Fondazione Berlucchi to the authors' laboratory is gratefully acknowledged

Correspondence

Angelo Manfredi H San Raffaele Institute DIBIT 3A1 via Olgettina 58 20132 Milano Italy Phone: +39-02.2643.4864 Fax: +39-02.2643.4786 E-mail address: manfredi.angelo@hsr.it

References

1. Okusaka T, Kosuge T. Systemic chemotherapy for pancreatic cancer. Pancreas 2004; 28:301-4. [PMID 15084976]

2. Rosenberg L, Lipsett M. Biotherapeutic approaches to pancreatic cancer. Expert Opin Biol Ther 2003; 3:319-37. [PMID 12662145]

3. Li D, Xie K, Wolff R, Abbruzzese JL. Pancreatic cancer. Lancet 2004; 363:1049-57. [PMID 15051286]

4. Rosenberg SA. Shedding light on immunotherapy for cancer. N Engl J Med 2004; 350:1461-3. [PMID 15070799]

5. Yu P, Lee Y, Liu W, Chin RK, Wang J, Wang Y, et al. Priming of naive T cells inside tumors leads to eradication of established tumors. Nat Immunol 2004; 5:141-9. [PMID 14704792]

6. Banchereau J, Steinman RM. Dendritic cells and the control of immunity. Nature 1998; 392:245-52. [PMID 9521319]

7. Lanzavecchia A, Sallusto F. Regulation of t cell immunity by dendritic cells. Cell 2001; 106:263-6. [PMID 11509174]

8. Vicari AP, Caux C, Trinchieri G. Tumour escape from immune surveillance through dendritic cell inactivation. Semin Cancer Biol 2002; 12:33-42. [PMID 11926410]

9. Bell D, Chomarat P, Broyles D, Netto G, Harb GM, Lebecque S, et al. In breast carcinoma tissue, immature dendritic cells reside within the tumor, whereas mature dendritic cells are located in peritumoral areas. J Exp Med 1999; 190:1417-26. [PMID 10562317]

10. Movassagh M, Spatz A, Davoust J, Lebecque S, Romero P, Pittet M, et al. Selective accumulation of mature DC-Lamp+ dendritic cells in tumor sites is associated with efficient T-cell-mediated antitumor response and control of metastatic dissemination in melanoma. Cancer Res 2004; 64:2192-8. [PMID 15026362] 11. Scaffidi P, Misteli T, Bianchi ME. Release of chromatin protein HMGB1 by necrotic cells triggers inflammation. Nature 2002; 418:191-5. [PMID 12110890]

12. Bianchi ME, Manfredi A. Chromatin and cell death. Biochim Biophys Acta 2004; 1677:181-6. [PMID 15020058]

13. Bellone M, Iezzi G, Rovere P, Galati G, Ronchetti A, Protti MP, et al. Processing of engulfed apoptotic bodies yields T cell epitopes. J Immunol 1997;159:5391-9. [PMID 9548479]

14. Rovere P, Vallinoto C, Bondanza A, Crosti MC, Rescigno M, Ricciardi-Castagnoli P, et al. Bystander apoptosis triggers dendritic cell maturation and antigen- presenting function. J Immunol 1998; 161:4467-71. [PMID 9794367]

15. Ronchetti A, Rovere P, Iezzi G, Galati G, Heltai S, Protti MP, et al. Immunogenicity of apoptotic cells in vivo: role of antigen load, antigen-presenting cells, and cytokines. J Immunol 1999; 163:130-6. [PMID 10384108]

16. Rovere P, Sabbadini MG, Vallinoto C, Fascio U, Zimmermann VS, Bondanza A, et al. Delayed clearance of apoptotic lymphoma cells allows crosspresentation of intracellular antigens by mature dendritic cells. J Leukoc Biol 1999; 66:345-9. [PMID 10449179]

17. Chiodoni C, Paglia P, Stoppacciaro A, Rodolfo M, Parenza M, Colombo MP. Dendritic cells infiltrating tumors cotransduced with granulocyte/macrophage colony-stimulating factor (GM-CSF) and CD40 ligand genes take up and present endogenous tumorassociated antigens, and prime naive mice for a cytotoxic T lymphocyte response. J Exp Med 1999; 190:125-33. [PMID 10429676]

18. Melero I, Vile RG, Colombo MP. Feeding dendritic cells with tumor antigens: self-service buffet or a la carte? Gene Ther 2000; 7:1167-70. [PMID 10918484]

19. Kurts C, Miller JF, Subramaniam RM, Carbone FR, Heath WR. Major histocompatibility complex class I-restricted cross-presentation is biased towards high dose antigens and those released during cellular destruction. J Exp Med 1998; 188:409-14. [PMID 9670054]

20. Srivastava PK, Menoret A, Basu S, Binder RJ, McQuade KL. Heat shock proteins come of age: primitive functions acquire new roles in an adaptive world. Immunity 1998; 8:657-65. [PMID 9655479]

21. Rovere-Querini P, Dumitriu IE. Corpse disposal after apoptosis. Apoptosis 2003; 8:469-79. [PMID 12975578]

22. Savill J, Dransfield I, Gregory C, Haslett C. A blast from the past: clearance of apoptotic cells

regulates immune responses. Nat Rev Immunol 2002; 2:965-75. [PMID 12461569]

23. Bondanza A, Zimmermann VS, Dell'Antonio G, Dal Cin E, Capobianco A, Sabbadini MG, et al. Cutting edge: dissociation between autoimmune response and clinical disease after vaccination with dendritic cells. J Immunol 2003; 170:24-7. [PMID 12496378]

24. Bondanza A, Zimmermann VS, Dell'Antonio G, Cin ED, Balestrieri G, Tincani A, et al. Requirement of dying cells and environmental adjuvants for the induction of autoimmunity. Arthritis Rheum 2004; 50:1549-60. [PMID 15146425]

25. Shi Y, Zheng W, Rock KL. Cell injury releases endogenous adjuvants that stimulate cytotoxic T cell responses. Proc Natl Acad Sci U S A 2000; 97:14590-5. [PMID 11106387]

26. Shi Y, Rock KL. Cell death releases endogenous adjuvants that selectively enhance immune surveillance of particulate antigens. Eur J Immunol 2002; 32:155-62. [PMID 11754356]

27. Shi Y, Evans JE, Rock KL. Molecular identification of a danger signal that alerts the immune system to dying cells. Nature 2003; 425:516-21. [PMID 14520412]

28. Massa C, Guiducci C, Arioli I, Parenza M, Colombo MP, Melani C. Enhanced efficacy of tumor cell vaccines transfected with secretable hsp70. Cancer Res 2004; 64:1502-8. [PMID 14973071]

29. den Brok MH, Sutmuller RP, van der Voort R, Bennink EJ, Figdor CG, Ruers TJ, Adema GJ. In situ tumor ablation creates an antigen source for the generation of antitumor immunity. Cancer Res 2004; 64:4024-9. [PMID 15173017]

30. Garcea G, Lloyd TD, Aylott C, Maddern G, Berry DP. The emergent role of focal liver ablation techniques in the treatment of primary and secondary liver tumours. Eur J Cancer 2003; 39:2150-64. [PMID 14522372]

31. de Jong KP, von Geusau BA, Rottier CA, Bijzet J, Limburg PC, de Vries EG, et al. Serum response of hepatocyte growth factor, insulin-like growth factor-I, interleukin-6, and acute phase proteins in patients with colorectal liver metastases treated with partial hepatectomy or cryosurgery. J Hepatol 2001; 34:422-7. [PMID 11322204]

32. Rovere P, Peri G, Fazzini F, Bottazzi B, Doni A, Bondanza A, et al. The long pentraxin PTX3 binds to apoptotic cells and regulates their clearance by antigenpresenting dendritic cells. Blood 2000; 96:4300-6. [PMID 11110705]

33. Gershov D, Kim S, Brot N, Elkon KB. C-Reactive protein binds to apoptotic cells, protects the cells from assembly of the terminal complement components, and sustains an antiinflammatory innate immune response:

implications for systemic autoimmunity. J Exp Med 2000; 192:1353-64. [PMID 11067883]

34. Manfredi AA, Iannacone M, D'Auria F, Rovere-Querini P. The disposal of dying cells in living tissues. Apoptosis 2002; 7:153-61. [PMID 11865199]

35. Nauta AJ, Daha MR, Kooten C, Roos A. Recognition and clearance of apoptotic cells: a role for complement and pentraxins. Trends Immunol 2003; 24:148-54. [PMID 12615211]

36. Muerkoster S, Wegehenkel K, Arlt A, Witt M, Sipos B, Kruse ML, et al. Tumor stroma interactions induce chemoresistance in pancreatic ductal carcinoma cells involving increased secretion and paracrine effects of nitric oxide and interleukin-1beta. Cancer Res 2004; 64:1331-7. [PMID 14973050]

37. Dallal RM, Christakos P, Lee K, Egawa S, Son YI, Lotze MT. Paucity of dendritic cells in pancreatic cancer. Surgery 2002; 131:135-8. [PMID 11854690]

38. Esche C, Lokshin A, Shurin GV, Gastman BR, Rabinowich H, Watkins SC, et al. Tumor's other immune targets: dendritic cells. J Leukoc Biol 1999; 66:336-44. [PMID 10449178]

39. Perrotta C, Falcone S, Capobianco A, Camporeale A, Sciorati C, De Palma C, Pisconti A, Rovere-Querini P, Bellone M, Manfredi AA, Clementi E. Nitric oxide confers therapeutic activity to dendritic cells in a mouse model of melanoma. Cancer Res 2004; 64:3767-71. [PMID 15172982]

40. Schmidt T, Ziske C, Marten A, Endres S, Tiemann K, Schmitz V, et al. Intratumoral immunization with tumor RNA-pulsed dendritic cells confers antitumor immunity in a C57BL/6 pancreatic murine tumor model. Cancer Res 2003; 63:8962-7. [PMID 14695214]

41. Kianmanesh A, Hackett NR, Lee JM, Kikuchi T, Korst RJ, Crystal RG. Intratumoral administration of low doses of an adenovirus vector encoding tumor necrosis factor alpha together with naive dendritic cells elicits significant suppression of tumor growth without toxicity. Hum Gene Ther 2001; 12:2035-49. [PMID 11747595]

42. Tong Y, Song W, Crystal RG. Combined intratumoral injection of bone marrow-derived dendritic cells and systemic chemotherapy to treat preexisting murine tumors. Cancer Res 2001; 61:7530-5. [PMID 11606390]

43. Teitz-Tennenbaum S, Li Q, Rynkiewicz S, Ito F, Davis MA, McGinn CJ, Chang AE. Radiotherapy potentiates the therapeutic efficacy of intratumoral dendritic cell administration. Cancer Res 2003; 63:8466-75. [PMID 14679011]

44. Furumoto K, Soares L, Engleman EG, Merad M. Induction of potent antitumor immunity by in situ

targeting of intratumoral DCs. J Clin Invest 2004; 113:774-83. [PMID 14991076]

45. Allavena P, Piemonti L, Longoni D, Bernasconi S, Stoppacciaro A, Ruco L, Mantovani A. IL-10 prevents the differentiation of monocytes to dendritic cells but promotes their maturation to macrophages. Eur J Immunol 1998; 28:359-69. [PMID 9485215]

46. Bondanza A, Rovere P, Borri A, Caremoli ER, Guidetti A, Citterio G, et al. Cytokine secretion associated with the clearance of apoptotic bodies in renal cell carcinoma patients. Int J Cancer 2001; 91:713-7. [PMID 11267985]

47. Vicari AP, Chiodoni C, Vaure C, Ait-Yahia S, Dercamp C, Matsos F, et al. Reversal of tumor-induced dendritic cell paralysis by CpG immunostimulatory oligonucleotide and anti-interleukin 10 receptor antibody. J Exp Med 2002; 196:541-9. [PMID 12186845]

48. Chan HH, Nishioka NS, Mino M, Lauwers GY, Puricelli WP, Collier KN, Brugge WR. EUS-guided photodynamic therapy of the pancreas: a pilot study. Gastrointest Endosc 2004; 59:95-9. [PMID 14722560]

49. Bhutani MS. Endoscopic ultrasound guided antitumor therapy. Endoscopy 2003; 35:S54-6. [PMID 12929056]