



Screening of Phytochemical Constituents of *Nymphaea Caerulea savigny*. An Aquatic Plant Resource for Drug Development

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ABSTRACT

Objectives: *Nymphaea caerulea* Savigny. is multipurpose medicinal plant has been mentioned for the treatment of liver disorders in Ayurveda, an ancient system of medicine. Hence the Phytochemical evaluation and estimation of secondary metabolites were carried out.

Methods: Polar and non-polar solvents were used to extract maximum number of compounds from various plant parts of *N. caerulea* preliminary screening analysis (Gibbs, 1974) and quantitative estimation of phytochemicals (Makkar *et al.*, 1993) was carried out.

Results: The results revealed that the plant is a rich source of different secondary metabolites like anthocyanins, anthraquinones, emodins, fatty acids, flavonoids, luecoanthocyanins, glycosides, phenols, coumarins, tannins and triterpenoids. Methanol, ethanol, chloroform and water extracts of leaf and flower are excellent source of phytocostituenpts when compare with rhizome and root.

Conclusion: The findings of the study will be helpful to the pharmacologists and phytochemists for identification of novel potential active compounds.

Keywords: Aquatic plant, Secondary metabolites, *Nymphaea caerulea*, Aqueous extract, Methanol extract, Ethanol extract and Chloroform extract.

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INTRODUCTION

The world is looking for new drugs to manage various diseases. All Ayurvada, Unani, Homeopathy and Siddha drugs are depending on herbals. Ayurveda is an ancient health care system and is practiced widely in India and other countries¹.

Roughly 20% of the plants found in the world have been submitted to pharmacological or biological test, and a substantial number of new antibiotics introduced in to the market are obtained from natural or semi-synthetic resources².

The investigation of phytochemical compounds like alkaloids, flavonoids, phenols, saponins, tannins, glycosides, terpenoids, coumarins etc. are highly essential to meet the pharmaceutical demand.

In India ten species of *Nymphaea*, both wild (*N. alba*, *N. candida*, *N. noucholii*, *N. pubescens*, *N. rubra* and *N. tetragona*) and cultivated (Fig.1 b) (*N. caerulea*, *N. marliacea*, *N. micrantha* and *N. alba* var *rubra*), are reported³. *Nymphaea* is globally distributed, cultivated, aquatic and Ayurvedic herb with perennial rhizomes or rootstocks anchored with mud with floating or submerged leaves and solitary and showy flowers⁴. It is used in Ayurvedic medicine mainly the flowers are used to wine dyspepsia, enteritis, diarrhoea, urinary problems, fevers and heart palpitations⁵. The leaves, roots, and flowers have a wide range of pharmacological activities and are used for diabetes, eruptive fevers and liver disorders⁶. Number of medicinal plants *Thespesia populnea* and *Tridax procumbens*⁷, underutilized species of Cyperaceae⁸. *Dysophylla myosuroides* and *Talinum cuneifolium*⁹ were evaluated for phytochemical screening. Though the plant is used in conventionally to cure several diseases, reports on evaluation of phytochemical constituents in *N. caerulea* (Fig.1 a) is not carried so far therefore, the present work is undertaken to screen and estimate the plant for secondary metabolites present in the root, rhizome, leaf and flower of *N. caerulea* as secondary metabolites present in plants are responsible for curing diseases.

MATERIALS AND METHODS

Roots, rhizomes, leaves and flowers (Fig.2 b, c, e and f) were collected from palakonda hills near palakondaraya temple, kadapa District. Washed the *N. caerulea* with tap water and was allowed to dry under

shade. The dried roots, rhizomes, leaves and flowers were made in to fine powders separately. These powders were extracted with water, ethanol and chloroform; then the preliminary photochemical tests were carried out for the detection of secondary metabolites.

1) Water extraction

10 g of air dried powder was added to 100 ml of distilled water and boiled for 2 h. The supernatant was collected and same procedure was repeated twice. The collected supernatant at an interval of every 2 h were pooled together and concentrated to make the final volume into one-fourth of the original volume. It was then autoclaved at 121°C and 15 lbs pressure and stored at 4°C.¹⁰

2) Chloroform, Ethanol and Methanol extraction

Dried roots, rhizomes, leaves and flowers each case, powdered air-dried plant materials were extracted with Chloroform, Ethanol and Methanol separately. The crude chloroform extract was prepared by maceration of plant materials (100 g each) with (300 ml) of each solvent¹¹; the extracts were filtered and stored in separate bottles.

Phytochemical screening

The condensed extracts were used for preliminary screening of phytochemicals such as flavonoids¹², steroids¹³, terpenoids¹⁴, tannins¹⁵, glycosides¹⁶, saponins¹⁴, alkaloids¹³, phenols¹³, quinones¹⁷, lignin¹³, coumarins¹⁸, leucoanthocyanins¹⁵ and emodins¹⁵.

Quantitative analysis of secondary metabolites

The rhizome, root, leaf and flower of *Nymphaea caerulea* were ground to pass through a sieve of 1 mm diameter. Tannins extraction was done using 400 mg ground

sample in conical flask with 40 ml diethyl ether containing 1 per cent acetic acid (v/v) and mixed to remove the pigment material and supernatant was carefully discarded after 5 min., 20 ml of 70 per cent aqueous acetone was added, sealed the flask with cotton plug covering with aluminum foil and kept in electrical shaker for 2 hrs for extraction. Then it was filtered through Whatman No.1 filter paper and sample was kept in refrigerator at 4° C until analysis. Then the estimation of phenols and tannins¹⁹, condensed tannins²⁰ and flavonoids²¹ were determined.

RESULTS AND DISCUSSION

The phytochemical screening of various parts of *N. caerulea* showed that they are rich in phenols, flavonoids, saponins, anthraquinones and anthocyanins present in all plant parts but leaves and flowers are showing all phytochemicals in methanol, ethanol and chloroform extractions. Leaves and flowers have more number of secondary metabolites than the rhizome and roots (Table-1). Triterpenoides, alkaloids are absent in root whereas leucoanthocyanins, anthraquinones, triterpenoides and alkaloids are absent in rhizome.

Alkaloids are found only in aqueous extract of flower and leaf. These are produced by large variety of organisms including bacteria, fungi, plants and animals; and are part of group of natural products; some alkaloids have a bitter taste while many are toxic to other organisms²². Anthocyanins are present in flowers, leaves and roots extracts of aqueous, chloroform, ethanol and methanol and absent in flowers, leaves, roots and rhizomes extracts of chloroform; and rhizome extracts of aqueous, ethanol, and methanol. Anthocyanins may help the human immune system to work more efficiently to protect against viral infections. It little bit more

complex, specific types of anthocyanins may have a direct effect in decreasing influenza viruses' infectivity by decreasing the ability of the virus itself to get into the human cell or to be related from infected cells or by having a viricide effect²². Anthraquinones are present in flower and leaf extract of aqueous; root extract of ethanol and flower, leaf and root extracts of methanol. These are absent in root and rhizome extracts of aqueous and flower, leaf, root and rhizome extracts of chloroform; flower, leaf and rhizome extract of ethanol and rhizome extracts of methanol. Anthraquinones are used for stomach-ache and in the treatment of diarrhoea²⁴ and these are an important chemical raw material and organic intermediates that are broadly applied in the field of dyestuff, papermaking, medicines and agricultural chemicals etc²⁵.

Coumarins are present only in flowers and leaves in all extracts. Various studies have been demonstrated that coumarins are potential antioxidants and their antioxidant activities are due to their ability to scavenge free radicals and to chelate metal ions²⁶. Emodins are found in flower and leaf extracts of aqueous and methanol. Emodins are one of the natural anticancer drugs, an active ingredient of the traditional Chinese medicine-rhubarb²⁷. Flavonoids are present in flower, leaf, root and rhizome extracts of aqueous, and absent in flower, root and rhizome extracts of chloroform, root and rhizome extract of ethanol and methanol. Flavonoids are a group of poly phenolic compounds influence the radical scavenging, inhibition of hydrolytic and oxidative enzymes and also act as anti-inflammatory agents²⁸, the flavonoids show antioxidant activity and their effects on human nutrition and health are considerable. The mechanisms of action of flavonoids are through scavenging or chelating process^{29, 30}. They also inhibit microbes which are resistance to

antibiotics³¹, flavonoids are free radical scavengers, super antioxidants and potent water soluble which prevent oxidative cell damage and have strong anti-cancer activity³², as an antioxidants, flavonoids provide anti-inflammatory activity³³.

Glycosides are present in flower and leaf extracts of aqueous, chloroform, ethanol and methanol; root extract of ethanol, rhizome extract of aqueous and ethanol. These are absent in root extract of aqueous, chloroform and methanol; rhizome extract of chloroform and methanol. Glycosides compounds are containing a carbohydrate and non-carbohydrates residues (moiety) in the same molecule. The carbohydrate moiety is attached an acetyl linkage carbon-I to the non-carbohydrate residue (aglycone). They all contain steroid as aglycone component in combination with sugar molecules. They are important in medicine because of their action on heart and are used in cardiac insufficiency³⁴. Leucoanthocyanins are present in flower and rhizome extracts of aqueous and ethanol; leaf and root extracts of methanol and absent in flower, leaf, root and rhizome extracts of chloroform; root and rhizome extracts of aqueous and ethanol, and flower and rhizome extracts of methanol. Leucoanthocyanins are occupying an important position among the water soluble organic compounds. They have been implicated as being responsible for the astringent taste of unripe fruit they are responsible for the chill haze that develops in beer and for the browning of white wine. They influence the storage stability of wine and juice³⁵. Steroids are present in flower and leaf extracts of aqueous, ethanol and methanol; flower extract of chloroform, rhizome extracts of aqueous, chloroform, ethanol and methanol and absent in leaf extract of chloroform, root extract of aqueous, chloroform, ethanol and methanol. It should be noted that steroids are of importance and of interest in pharmacy due

to their relationship with sex hormones³⁶. Phenols are present in flower and leaf extracts of aqueous, ethanol and methanol, leaf extract of chloroform, root and rhizome extracts of aqueous and absent in flower extract of chloroform, root and rhizome extracts of chloroform, ethanol and methanol. Bioactive polyphenols have attracted special attention because they can protect the human body from the oxidative stress which may cause many diseases, including cancer, cardiovascular problems and ageing³⁷. Many of the phenols have shown to contain higher levels of anti oxidants activities^{38,39}, have reported that plant phenols in red wine exerted cardio protective effect.

Saponins are found in flower, leaf, root and rhizome extracts of aqueous; flower, leaf and rhizome extracts of chloroform; flower and rhizome extracts of ethanol, leaf and root extracts of methanol and absent in root extract of chloroform and ethanol; leaf extract of ethanol, flower and rhizome extracts of methanol. Saponins are used as detergents, pesticides and molluscicides, in addition to their industrial application as foaming and surface active agents and also have beneficial health effects⁴⁰. Tannins are present in flower, leaf, root and rhizome extracts of aqueous; leaves extract of chloroform; flower and leaf extracts of ethanol and methanol and absent in flower, root and rhizome extracts of chloroform, root and rhizome extract of ethanol and methanol. Tannins contribute property of astringency i.e. fasten the healing of wound and inflamed mucous membrane and have received considerable attention in the fields of nutrition, health and anti inflammatory properties⁴¹. Tannins are complex moieties produced by majority of plants as protective substances; they possess wide pharmacological activities and have been used since past as tanning agents and they posses astringent, anti diarrhoeal and

anti-inflammatory activities⁴². Triterpenoids are present in flower and leaf extracts of aqueous, chloroform and methanol. Triterpenoids are absent in flower and leaf extracts of ethanol, and root and rhizome extracts of aqueous, chloroform, ethanol and methanol. Triterpenoids are attributed for analgesic and anti-inflammatory activities. The results revealed that the saponins and glycosides are found in all extracts used in present work. Aqueous extract is the best among the solvents used for extraction of secondary metabolites. Flowers and leaves are rich source of phytochemical constants in order.

Based on the screening of secondary metabolites we conducted the quantitative estimation of some important secondary metabolite like flavonoid, phenols and tannins. The highest total flavonoid content was found in flowers (0.02275 mg QE/g (Quercetin Equivalent)/g) followed by leaf (0.012 QE/g) whereas absent in rhizome and root. Flavonoids are reported to possess many useful properties, including anti-inflammatory, antimicrobial, enzyme inhibition, oestrogenic, antiallergic, antioxidant and antitumour activity^{43, 44}.

The highest total phenols content was found in leaf (325 mg TAE (Tannic Acid Equivalent)/g) than flower (315 mg TAE/g), rhizome (210 mg TAE/g) and root (1801 mg TAE/g) (Table-2). Similar results have been shown in *Pongamia* which contain higher levels of antioxidant activity⁴⁵. The higher amount of phenols is important in the regulation of plant growth, development and diseases resistance. It can be used as fungicide, pesticides, an antiseptic, disinfectant and in the manufacture of resins, explosives, plastics, detergents and pharmaceutical substances⁴⁶, *t*⁴⁷.

The highest total tannins content was found in the leaf (90.156 mg TAE/g) and followed by flower (0.11225 mg TAE/g).

The growth of many fungi, yeasts, bacteria and viruses was inhibited by tannins. Apart from these tannins contribute the property of astringent activity i.e. faster the healing of wounds and inflamed mucous membrane⁴⁸. Studies have indicated that antioxidants prevent the onset of degenerative illness such as certain cancers, cardiovascular and neurodegradative diseases⁴⁹.

CONCLUSION

Nymphaea caerulea plants possesses number of secondary metabolites particularly rich source of saponins, glycosides and steroids. The plant widely had used in traditional medicine to combat and cure various ailments. The anti-inflammatory, anti-spasmodic, anti-cancer, anti-microbial, anti-analgesic, anti-oxidants and anti-diuretic activities can be attributed due to the presence of higher levels of steroids, tannins, terpenoids, flavonoids, phenols, emodins and saponins. Exploitation of these pharmacological properties involved further investigation of these active ingredients by implementation of techniques of extraction, purification, separation, crystallization and identification.

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REFERENCES

- Chopra, A. and Doiphode, V.V. (2002) 'Ayurvedic medicine. Core concept, therapeutic principles, and current relevance, Medical Clinics of North America, Vol. 86, pp. 75-89.
- Mothana, R. and Lindequist, U. (2005). Antimicrobial activity of some medicinal plants of the island Socotra *J. Ethnopharmacol.* 96:177-181.

3. Mitra RL (1990). Nymphaeaceae. In: Nayar MP, Thothathri K, Sanjappa M (eds.) *Fascicles of flora of India, Fascicles 20. Botanical Survey of India, Kolkata.* 11–25.
4. Conard, H.S., 1905. The water lilies: a monograph of the genus *Nymphaea*. Carnegie inst. Wash. Publ. 4, 1-279.
5. Encyclopedia of Herbs and their uses 1995. Devi Bown. Dorling Kindersley Limited, London, New York, Stuttgart, Moscow, p. 317.
6. Dhanabal SP, MohanMarugaRaja MK, Ramanathan M, SureshB, (2007). Hypoglycemic activityof *Nymphaea stellata* leaves ethanoliceextractinalloxan induced diabeticrats. *Fitoterapia.* **78:** 288–291.
7. Savithramma N, Linga Rao M and Bhumi G (2011). Phytochemical screening of *Thespesia populnea* (L.) Soland and *Tridax procumbens*. *L J Chem Pharm Res.* **3(5):** 28-34.
8. Hari Babu R and Savithramma N (2014). Screening of Secondary Metabolites of Underutilized Species of Cyperaceae. *Int J Pharm Sci Rev Res.* **24(2):** 182-187.
9. N. Savithramma and M. Linga Rao and Beena Prabha, 2011. Phytochemical Studies of *Dysophylla yosuroides* (Roth.) Benth. In. Wall. And *Talinum cuneifolium* (Vahl.) Willd. *Research Journal of Phytochemistry,* **5:** 163-169.
10. Parekh Jigna, sumitra CV (2007). Invitro antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turk J Biol.* **31:** 53-58.
11. Alanis AD, Calzada F and Caervantes JA (2005). Antimicrobial properties of some plants used in Mexican Traditional Medicine for the treatment of gastrointestinal disorders. *Ethnopharmacology.* **100:** 153-157.
12. Peach K and Tracey MV (1956). Modern methods of plant analysis. *Springer Verlag, Berlin.* **3:**
13. Gibbs RD, Chemotaxonomy of Flowering Plants, Mc Gill Queen's University Press, Montreal and London, 1, 1974.
14. Ayoola GA, Coker HAB, Adesegun SA, Adepoju-Bello AA, Obaweya K, Ezennia EC and Atangbayila TO (2008). Phytochemical plants used for malaria therapy in South Western Nigeria. *Trop J Pharm Res.* **7:** 1019-1024.
15. Treare GE and Evans WC (1985). Pharmacognosy, Bahive Tinal, London. **17:** 149.
16. Kokate CK, Purohit AP and Gokhale SB (1999). Pharmacognosy, *Nirali publishers, Pune,* 1-2.
17. ASEAN countries (1993). Standard of ASEAN herbal medicine, *Jakatra Buena Printing.* **1:** 116-28.
18. Rizk AM, Constituents of plant growing in Qatar. I. A chemical survey of sixty plants. *Fitoterapia,* (1982); 52-42.
19. Makkar HPS, Blummel M, Borowy NK and Becker K (1993). Gravimetric determination of tannins and their correlations with chemical and protein precipitation methods. *J Sci Food Agri* **61:** 161-165.
20. Porter LJ, Hrstich LN and Chan BJ (1986). The conversion of proanthocyanidin prodelphinidins to cyanids and delphinidin. *Phytochemistry* **25:** 223-230.
21. Chang C, Yang M Wen H and Chern J (2002). Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *J Food Drug Analysis* **10:** 178-182.
22. Gupta VK, Singh GD, Singh S and Kaul A (2010). Medicinal plants: Phytochemistry, Pharmacology and Therapeutics. **Daya Publishing House, Delhi.**
23. Liu et al., (2009) in vitro anti-influenza viral activities of constuens from

- Caesalpinia sappan. *Planta Med.* **100**:3-13.
24. Sabinis SD and Daniel M (1990). A phytochemical approach to economic Botany, *Kalyani Publishers, New Delhi.* **15**: 65.
25. Anthraquinons information available from www.sheri.com
26. Tseng A Chemoprevention of tumors in MTV-H ras transgenic mice with coumarins. *Proc. Am. Assoc. Cancer. Res.,* (1991); **32**:2257.
27. Khemkaran A and Jain SK (2011). Aloë-emodin novel anticancer herbal drug. *In J Phytomed.* **3**: 27-31.
28. Frankel E (1995). Nutritional benefits of flavonoids. International conference on food factors: chemistry and cancer prevention, hamamatsu. *Japan Abstracts.* **6**: 2.
29. Kessler M, Ubeand G and Jung L (2003); Anti and prooxidant activity of rutin and quercetin derivatives. *J Pharm and Pharmacol.* **55**: 131-142.
30. Cook NC and Samman S (1996). Flavonoids chemistry, metabolism, cardioprotective effects, and dietary sources. *Nutri Bioche.* **7**: 66- 76.
31. Linuma M, Tsuchiya H, Sato M, Yokoyama J, Ohyama M, Ohkawa Y, Tanaka T, Fujiwara S and Fujii T (1994). Flavanones with potent antibacterial activity against methicillin-resistant *Staphylococcus aureus*. *J Pharmacol.* **46** (11): 892-895.
32. Salah W, Miller N, Pagauga G, Tybury G, Bolwell E, Rice E and Evans C (1995). Polyphenolic flavonoids as scavenger of aqueous phase radicals and chain breaking antioxidants. *Arch Biochem.* **2**: 239-346.
33. Okwu DE (2001). Improving the nutrition value of *Cassava tapioca* meal with local species. *Nutraceutical, Functional and medicinal food.* **3**: 43-51.
34. Balch JF and Balch PA (2000). Prescription for Nutritional Healing. *New York. A very,penguin Putnam Inc.* 267-270.
35. Josly MA and Goldstein L (1953). Astringency of fruit and fruit products in relation to leucoanthocyanin content. Progress report of Agricultural research, The University of California Division of Agricultural Science.
36. Santhi R, Lakshmi G, Priyadarshini AM and Anandaraj L (2011). Phytochemical screening of *Nerium oleander* leaves and *Momordica charantia* leaves. *Inter Res J Pharm.* **2**:131-135.
37. Robards K, Prernzler PD, Tucker G, Swatsitang P and Glover W (1999). Phenolic compounds and their role in oxidative processes in fruits. *Food Chem.* **66**: 401-36.
38. Razali N, Razab R, Mat Junit S and Abdul Aziz A. (2008). Radical scavenging and reducing properties of extracts of cashew shoots (*Anacardium occidentale*). *Food Chem.* **111**: 38-44.
39. Fogliano V, Verde V, Randazzo G and Ritienti A (1999). Method of measuring antioxidant activity and its application to monitoring the antioxidant capacity of wines. *J Agric Food Chem.* **47**:1035-1040
40. Arunasalam Shi J, Yeung K, Kakuda D, Mittal Y and Jiang Y (2004). saponins from edible legumes: chemistry, processing and health benefits. *J Med Food.* **7**: 67-78.
41. Santos-Buelga C and Scalbert A (2000). Proanthocyanidins and tannin like compounds nature, occurrence, dietary intake, and effects on nutrition and health. *J Sci Food Agric.* **80**: 1094-1117.
42. Killedar SG and More HN (2010). Estimation of tannins in different parts of *Memecylonum bellatum*. *Burm J Phar Res.* **3**(3): 554-556.

43. Havesteen B (1990). Flavonoids a class of natural products for antimicrobial agents. *Eur J Clin Microbial Infect Dis*, 9: 455-61.
44. Harborne JB and Williams CA (2000). Advance in flavonoid research since 1992. *Phytochemistry*. 55: 481-504.
45. Shirwaikar A, Malini S and Kumari SC (2003). Protective effect of *Pongamia pinnata* flowers against cisplatin and gentamicin induced nephrotoxicity in rats. *Indian J Exp Biol*. 1: 58-62.
46. Savita Sagwan, Rao DV and Sharma RA (2010). biochemical estimation of primary metabolites from pongamia pinnata (l.): an important biodiesel plant. *Inter J Pharm Sci Rev Res*. 5 (1): 146-149.
47. Amani M D El-Mesallamy, Fatma A Ahmed, Mohammed H Elhaw and Taha A Ibrahim (2015). Chemical investigation and evaluation of antimicrobial activity of *trichodesma ehrenbergii* schweinf. Ex boiss. growing widely in gebel elba, Egypt. *Indo American J of Pharm Sci*, 2(5): 961-966.
48. Cheng KT, Wong TY, Wei CL, Huang YW and Lin Y (1998). Tannins and human health: A review. *Criti Rev Food Sci Nutri*. 6: 421-64.
49. Okwu DE, and Josiah C (2006). Evaluation of the chemical composition of two Nigerian medicinal plants. *Afr J Biotech*. 5: 357- 361.

Table 1. Qualitative analysis of phytochemical constituents of various parts of *Nymphaea caeruleae*.

| S. No | Phytochemical constituents | Aqueous | | | | Chloroform | | | | Ethanol | | | | Methanol | | | |
|----------|-------------------------------|---------|---|---|----|------------|---|---|----|---------|---|---|----|----------|---|---|----|
| | | F | L | R | Rh | F | L | R | Rh | F | L | R | Rh | F | L | R | Rh |
| 1. | Alkaloids | + | + | - | - | - | - | - | - | - | - | - | - | - | - | - | - |
| 2. | Anthocyanins | + | + | + | - | - | - | - | - | + | + | + | - | + | + | + | - |
| 3. | Anthroquinones | + | + | - | - | - | - | - | - | - | - | + | - | + | + | + | - |
| 4. | Coumarins | + | + | - | - | + | + | - | - | + | + | - | - | + | + | - | - |
| 5. | Emodins | + | + | + | - | - | + | - | - | - | - | - | - | + | + | - | + |
| 6. | Flavonoids | + | + | + | + | - | + | - | - | + | + | - | - | + | + | - | - |
| 7. | Glycosids | + | + | - | + | + | + | - | - | + | + | + | + | + | + | - | - |
| 8. | Leucoanthocyanins | + | + | - | - | - | - | - | - | + | + | - | - | + | + | - | - |
| 9. | Steroids | + | + | - | + | + | - | - | - | + | + | - | + | + | + | - | + |
| 10. | Phenols | + | + | + | + | - | + | - | - | + | + | - | - | + | + | - | - |
| 11. | Saponins | + | + | + | + | + | + | - | + | + | - | - | + | - | + | + | - |
| 12. | Tannins | + | + | + | + | - | + | - | - | + | + | - | - | + | + | - | - |
| 13. | Triterpenoids | + | + | - | - | + | + | - | - | - | - | - | - | + | + | - | - |

Note: '+' indicates presence, ' - ' indicates absence

(F) Flower, L) Leaf, R) Root and Rh) Rhizome)

Table 2. Quantitative estimation of secondary metabolites

| S NO. | <i>N. caerulea</i> | Flavonoids Mg QE/g | Phenols mg TAE/g | Tannins mg TAE/g |
|-------|--------------------|-----------------------|---------------------|---------------------|
| 1 | Flower | 0.02275 | 315 | 011225 |
| 2 | Leaf | 0.012 | 325 | 0.156 |
| 3 | Rhizome | - | 210 | - |
| 4 | Root | - | 180 | - |

‘_’ indicates not found/nil



Fig. 1 a) Author with plant b) Natural habitata of *N. caerulea*



Fig. 2 Various parts of *N. caerulea*
a) Plant b) Roots c) Rhizome d) T.S. of Rhizome e) Leaf f) Flower