

REVIEW

ICAM-1 and Acute Pancreatitis Complicated by Acute Lung Injury

XiPing Zhang¹, Dijiong Wu², Xinge Jiang²

¹Department of General Surgery, Hangzhou First People's Hospital;

²Zhejiang University of Traditional Chinese Medicine. Hangzhou, Zhejiang Province, China

Summary

One of the most common complications of acute pancreatitis is acute lung injury, during which intercellular adhesion molecule-1 (ICAM-1) plays an important role by participating in leukocyte adhesion and activation as well as by inducing the "cascade effect" of inflammatory mediators, pulmonary microcirculation dysfunction and even acute respiratory distress syndrome, multiple organ failure or death. Although it is generally believed that the modulatory mechanism of ICAM-1 during this process is associated with the activation of nuclear transcription factor kappa B which is mediated by IL-1, IL-6, IL-18 and oxygen free radical, etc., further studies are still required to clarify it. Since the upregulation of ICAM-1 expression in the lung during acute lung injury is one of main pathogeneses, the early detection of the ICAM-1 expression level may contribute to the prevention and treatment of acute lung injury. Moreover, reducing pulmonary ICAM-1 expression levels through treatment with anti-ICAM-1 monoclonal antibody (aICAM-1) and antagonists of the neurokinin 1 receptor, etc., should have a positive effect on protecting the lungs during acute pancreatitis. This review aims to further clarify the relationship between ICAM-1 and acute pancreatitis complicated by acute lung injury, and therefore provides a theoretical basis for the formulation of corresponding therapeutic measures in clinical practice for acute pancreatitis.

Introduction

Severe acute pancreatitis is a commonly encountered and frequently-occurring disease, which does harm to the pancreas as well as other organs. Multiple organ dysfunction syndromes and multiple organ failure are believed to be the main reasons for high mortality (11.8-25% [1, 2]) in severe acute pancreatitis. In particular, acute lung injury and acute respiratory distress syndrome are the most common complications during the first week after the onset of acute pancreatitis and mortality can reach 60% [3].

At present, effective therapeutic methods on preventing and treating acute pancreatitis complicated by acute lung injury are still vague, and the pathogenesis is not clear yet. Some reports have indicated that intercellular adhesion molecule-1 (ICAM-1) plays an important role

in the development and progression of acute pancreatitis complicated by acute lung injury, and the severity of the lung injury correlates well with the expression levels of ICAM-1 protein [4, 5, 6]. In this paper, we review the advances in research in this field.

1. Overview of ICAM-1

ICAM-1, a single-chain transmembrane glycoprotein with a molecular weight of 80-110 KDa, consists of five Ig-like domains, a hydrophobic transmembrane domain and a short cytoplasmic C-terminal domain [7]. Its ligand includes lymphocyte function-associated antigen-1 (LFA-1) and macrophage antigen-1 (Mac-1) [8]. Under physiological conditions, ICAM-1 is expressed at a low level in endothelial cells and epithelial cells [9, 10] or constitutively on the surface of alveolar cells [11], providing the underlying molecular basis for cell recognition, activation, proliferation, differentiation and motility, and thereby helping to stabilize the internal environment of the body. Moreover, ICAM-1 also plays a key role during pathological conditions, such as inflammatory reaction etc. [12, 13]. For these reasons, a comprehensive and objective understanding of ICAM-1 is needed.

2. ICAM-1 Expression and the Factors Influencing ICAM-1 Expression

2.1 Mechanism of ICAM-1 Expression

Experimental studies concerning acute pancreatitis have shown that an infusion of trypsin into the lungs

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Abbreviations LFA-1: lymphocyte function-associated antigen-1; Mac-1: macrophage antigen-1; NK1R: neurokinin-1 receptor; PPAR: peroxisome proliferator-activated receptors-gamma

Correspondence XiPing Zhang
Department of General Surgery, Hangzhou First People's Hospital,
Hangzhou 310006, Zhejiang Province, China
Phone: +86-571.8706.5701; Fax: +86-571.8791.4773
E-mail: zxp99688@vip.163.com

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can significantly upregulate the expressions of both membrane-bound ICAM-1 (mICAM-1) and soluble ICAM-1 (sICAM-1) proteins [14], and the use of a urinary trypsin inhibitor is obviously capable of downregulating the expression of ICAM-1 [15], suggesting that trypsin is an initial provocative factor for the expression of ICAM-1. At present, it is generally believed that the upregulation of ICAM-1 expression is associated with enhanced transcription of the ICAM-1 gene which might be activated by nuclear transcription factor kappa B (NF-kappa B) [16], although the precise mechanism is not entirely clear. NF-kappa B is a member of the Rel protein family and is maintained in the cytoplasm as an inactivated trimer complex through binding to the inhibitory protein I-kappa B under normal circumstances [17]. When acute pancreatitis complicated with acute lung injury occurs, inflammatory mediators gathering in the lungs can induce, through signal transduction systems, the phosphorylation of I-kappa B and its dissociation from NF-kappa B which subsequently translocates into the nucleus, binds to the promoter of the ICAM-1 gene and stimulates its transcription [18].

2.2 Influencing Factors of ICAM-1 Expression

The “cascade effect” of inflammatory mediators during acute pancreatitis could lead to the release of a large number of inflammatory mediators and various active substances into the blood, which subsequently enter into the lungs through the circulatory system and thereby result in lung injury and regulate the expression of ICAM-1. The main influencing factors of ICAM-1 expression include the following.

2.2.1 Tumor Necrosis Factor-alpha (TNF-alpha)

TNF-alpha, mainly produced by mononuclear cells, is not only capable of directly killing cells but is also capable of promoting the production of other cytokines [19], thus participating in the systemic progression of diseases from local stages. The content of TNF-alpha in serum increases significantly when acute pancreatitis complicated by acute lung injury occurs [20, 21]. For this reason, serum TNF-alpha concentration is considered to be an early parameter for reflecting the severity of acute pancreatitis. Some experimental results have shown that the binding of highly expressed TNF-alpha to TNF receptor (TNFR) is able to promote NF-kappa B-mediated ICAM-1 expression [22, 23], and the activation of NF-kappa B can in turn enhance the transcription of TNF-alpha gene, thereby forming a positive feedback loop which is able to amplify the early inflammatory signal and aggravate the initial inflammatory effect. In contrast, the soluble TNF receptor (sTNFR), as one of the ways of removing the TNF receptor [24], is able to inhibit the response of cells to TNF-alpha, thereby exerting a negative regulatory role and, to a certain extent, antagonizing ICAM-1-mediated lung injury.

2.2.2 Interleukin (IL) Family

Interleukins are important cytokines which play important roles during acute pancreatitis complicated by acute lung injury by means of stimulating the activation of leukocytes and the release of toxic substances, etc. [25, 26]. It has been proven that the expression levels of IL-1 and IL-6 have a good correlation with the expression levels of ICAM-1 in the lungs during acute pancreatitis [27, 28, 29]. Recently, some researchers have found that the expression levels of IL-18 are significantly elevated in patients with acute pancreatitis; IL-18 can promote the expression of ICAM-1 [30, 31, 32]; there exists a mutual induction between the expression of IL-18 and those of IL-1 and TNF-alpha, and IL-18 is involved in the functional failure of distant organs. The mechanism with which IL-18 exerts its functions must still be elucidated. Furthermore, IL-10 is considered to be capable of inhibiting the synthesis of a wide variety of pro-inflammatory cytokines [33, 34] and of perhaps having a negative regulatory effect on ICAM-1. In short, ILs play a dual role in the regulation of ICAM-1. During acute pancreatitis, persistent inflammatory stimulation enables the positive feedback to be dominant and, therefore, the injurious effect is more obvious.

2.2.3 Other Influencing Factors

Some protein kinase C (PKC) isoforms, such as effectors of G-protein-coupled receptor systems, are also involved in NF-kappa B activation upon receiving extracellular stimuli [35]. Some studies have indicated that this activation is associated with mitochondrial reactive oxygen species (mROS) which are regulated by protein kinase C isoforms [36], and Ca²⁺ plays a key role during this process. Therefore, calcium antagonists should be effective in reducing ICAM-1 expression. Furthermore, platelet-derived growth factor (PDGF), vascular endothelial growth factor (VEGF) and cyclooxygenase-2 (COX-2) are also considered to be directly related to the development of acute pancreatitis complicated by acute lung injury [37]. During these processes, the enhancement of NF-kappa B activity is the key point [22, 38, 39] since NF-kappa B can be synergistic with ICAM-1 expression in the lungs or on the surface of endothelial cells. As we know, reactive oxygen species (ROS) can promote the expression of ICAM-1 as well. The activated polymorphonuclear (PMN) leukocytes may be an important source of ROS which contributes to the massive expression of ICAM-1 on the surface of endothelial cells [40]. Interferon-gamma (INF-gamma) is one of the regulators of inflammation, and its upregulated expression is noted during respiratory distress syndrome complicating acute pancreatitis [41, 42]. It is generally believed that interferon-gamma is relevant to the development of acute pancreatitis. However, a recent study shows that inflammatory reaction in mice

with defects of INF-gamma expression is more severe than that in wild-type mice, suggesting that INF-gamma may function by means of suppressing the activity of NF-kappa B [43], although this result still needs to be verified.

3. Roles of ICAM-1 During Acute Pancreatitis Complicated by Acute Lung Injury

3.1 Inducing the Emigration and Recruitment of Leukocytes

The recruitment of leukocytes (mainly PMN) in the lungs plays a central role during acute pancreatitis complicated by acute lung injury [44]. A variety of adhesion molecules are involved in the adhesion to endothelial cells, migration through vascular endothelium and gathering in the lungs of PMN leukocytes [45, 46]. When PMN leukocytes are activated by IL-1, platelet-activating factor (PAF) or mast cells [47, 48], the expression of LFA-1 on the surface of PMN leukocytes is enhanced. The configuration change of the extracellular receptor-binding domain of LFA-1 is the key to the stable adhesion of PMN leukocytes. Cascade reactions induced by LFA-1 through binding to ICAM-1 can cause the structural rearrangement of PMN leukocytes, which subsequently migrate to inflammation sites in the presence of chemotactic factors and are enriched there [49]. In contrast, this phenomenon is not pronounced in mice with defects of ICAM-1 expression [50]. These results indicate that it is of great significance for ICAM-1 to mediate the adhesion and recruitment of PMN leukocytes, suggesting that ICAM-1 can be chosen as a target for treatment. Through blocking the binding of ICAM-1 to its ligands, it is expected that a better interference to the adhesion and migration of leukocytes would be achieved.

3.2 Promoting the Activation of Leukocytes

After PMN leukocytes adhere and bind to target cells, a relatively compact and stable micro-environment is formed, thus providing a necessary condition for toxic mediators to exert toxic action against target cells. The binding of ICAM-1 to Mac-1 and LFA-1 can further induce the expression of the latter on the surface of PMN leukocytes, and thereby promote PMN leukocytes to release hydrogen peroxide (H₂O₂) and a variety of pro-inflammatory cytokines [51, 52, 53]. Once inflammatory injury occurs, pulmonary epithelial cells and capillary endothelial cells will release a large number of blood coagulation-promoting substances which can cause platelet aggregation [54], and then promote an increase of pulmonary vascular resistance and permeability as well as interstitial pulmonary edema [55] and acute lung injury. These results demonstrate that the injuries of the lungs and other organs during acute pancreatitis are due to the activation of leukocytes. Although ICAM-1 is involved in the activation of PMN leukocytes, it is not the trigger factor. Therefore, inhibiting the expression of ICAM-1 is not capable of completely blocking the

activation of PMN leukocytes. Nevertheless, this inhibition may exert a positive effect in protecting lung injury caused by PMN leukocytes and delay the progression of the illness.

4. Expression Levels of ICAM-1 and the Diagnosis and Treatment of Acute Pancreatitis Complicated by Acute Lung Injury

4.1 Diagnostic Significance

Due to the particular physiological structure of the lungs, they are usually the first target of attack when multi-organ dysfunction complicating acute pancreatitis occurs. It is reported that about 48.3% of acute pancreatitis patients suffer from pulmonary disease, and part of them die from acute lung injury [56]. Therefore, it is of great practical significance to better predict acute lung injury and reduce its incidence rate. Through detecting the changes of serum ICAM-1 contents in 34 cases of patients with acute pancreatitis, some researchers have hypothesized that ICAM-1 can be used as an early marker in the diagnosis of lung injury [57]. If corresponding therapeutic measures are taken as soon as possible on the basis of early diagnosis through detecting ICAM-1, the therapeutic effect will certainly be superior to that achieved by conventional treatment. Currently, some researchers have attempted to conduct clinical studies on the classification of acute pancreatitis by standardizing the expression levels of ICAM-1 [58] since they believe that the extent and manner of the increase in blood sICAM-1 content in patients with acute pancreatitis are correlated with the severity of the disease. Although we also believe that the expression levels of ICAM-1 have significance in the classification of acute pancreatitis complicated by acute lung injury, further studies are still needed to verify this.

4.2 Therapeutic thoughts and Approaches

Since the precise mechanism underlying the development of acute pancreatitis complicated by acute lung injury has not yet been completely clarified, supportive therapy (for example, application of antibacterial drugs or nutritional support etc.) is generally used in the treatment of acute lung injury except when clear indications for surgery are clinically observed. However, these therapeutic measures are not able to fundamentally solve all the problems. Therefore, the exploration of new therapeutic strategies and approaches has become one of the focal points of the study. Some scholars have found that abnormal expressions of adhesion molecules, such as ICAM-1, during acute pancreatitis occur after the appearance of cytokines, and neutrophil infiltration and organ injury often occur after the upregulation of ICAM-1 expression. Therefore, they think that cell adhesion can be blocked by treatment [59]. For these reasons, studies on adhesion molecules have become a new breakthrough point. At present, many therapeutic approaches aimed at changing the expression levels of ICAM-1 have been proposed, including the following:

4.2.1 Treatment with Monoclonal Antibodies

The literature has shown that treatment using membrane-bound ICAM-1 as a target can effectively reduce lung injury [60], and better therapeutic effects can be achieved by using anti-ICAM-1 monoclonal antibody (aICAM-1) in the treatment of various types of acute lung injury in animal models [61]. If aICAM-1 is applied in the early stage of acute pancreatitis, better protection for the pancreas and lung functions is observed [62]. The underlying mechanism may be mainly associated with the inhibition of ICAM-1-mediated adhesion and infiltration of PMN leukocytes as well as the decrease in the expression of active substances, such as inflammatory mediators, etc. In addition, aICAM-1 is also able to lessen myeloperoxidase (MPO) and downregulate the expression of nitric oxide synthase-2 mRNA (NOS-2 mRNA) [63], thus ensuring the functional and structural stability of endothelial cells and exerting a positive influence on the protection of pulmonary microcirculation. However, it should also be noted that the long-term use of ICAM-1 antibodies may give rise to autoimmune diseases. Moreover, considering that this practice can only mitigate the disease, the popularization of ICAM-1 antibodies is still an issue open to question.

4.2.2 Treatment by Inhibiting NF-kappa B Activation

It is obvious that NF-kappa B plays a critical role in the expression of ICAM-1. Therefore, research on the use of NF-kappa B inhibitor to alleviate inflammation response has become a hotspot [64, 65, 66]. Calpain I inhibitor and pyrrolidine dithiocarbamate (PDTC) are antioxidants which are potent inhibitors of NF-kappa B. Calpain I inhibitor and pyrrolidine dithiocarbamate (PDTC) can lessen lung injury in rats with acute pancreatitis, decrease the activation of NF-kappa B as well as the expression of ICAM-1 protein [67, 68, 69], and can retain the soakage of inflammatory cells and mitigate the microvascular impairment of the lungs, which reduces the incidence rate of pneumonodema [70]. After Hietaranta *et al.* [71] first reported that MG132, a proteasome inhibitor, could depress the activation of NF-kappa B in acute pancreatitis, some researchers demonstrated that MG132 also had the effect of protecting lung tissue in rats with acute pancreatitis [72, 73, 74] which may be associated with the function of inhibiting NF-kappa B activation. The use of the NF-kappa B inhibitor may be considered as another effective path in the treatment of acute pancreatitis complicated by acute lung injury, and associated clinical research is required.

4.2.3 Treatment Through Activating Peroxisome Proliferator-Activated Receptors-gamma (PPAR-gamma)

A large number of studies have shown that, for acute pancreatitis or for chronic pancreatitis, the use of thiazolidinediones, pioglitazone and rosiglitazone, etc. [75, 76, 77] can antagonize the expression of TNF-alpha and ICAM-1, and delay the progression of

acute pancreatitis. Utilizing PPAR-gamma gene knockout mice, some researchers have proven that the functions of the above-mentioned drugs are mediated by PPAR-gamma [78]. Therefore, it is believed that PPAR-gamma agonists are able to effectively protect the lungs and reduce the expression of the ICAM-1 protein. The action mechanism of PPAR-gamma agonists is also associated with suppressing the activities of transcription factors NF-kappa B and activator protein-1 [79, 80, 81]. At present, the study of PPAR-gamma agonists in the treatment of acute pancreatitis complicated by acute lung injury is quite limited, although it has good prospects.

4.2.4 Other Therapeutic Strategies and Approaches

Neuropeptides, such as substance P (SP), can stimulate elevated expression of ICAM-1 in human skin capillary endothelial cells [82]. When neurokinin-1 receptor (NK1R) antagonists are used to treat acute pancreatitis mice, RT-PCR results reveal that the expression levels of ICAM-1 mRNA in the lungs in the study group are significantly lower than those in the model control group. Moreover, pathological changes in mice in the study group are also milder than those in the model control group [83]. Although the mechanism by which NK1R antagonists act on acute pancreatitis complicated by acute lung injury is not entirely clear, what is certain is that this practice is less likely to inhibit the activation of ICAM-1 and NF-kappa B. It has been proven that NF-kappa B activation happens prior to the transcription of ICAM-1 and NK1R mRNAs [84]. Thus, it is difficult to achieve a therapeutic effect on acute lung injury by antagonizing NK1R.

Conclusions

To sum up, ICAM-1 plays an extremely important role during the whole development process of acute pancreatitis complicated by acute lung injury, including the adhesion and activation of leukocytes and pulmonary vascular endothelial cells. The usage of aICAM-1, PPAR-gamma agonists, NF-kappa B inhibitor and antagonists of neurokinin 1 receptor, etc. should have a positive effect on reducing the expression of ICAM-1 in lungs during acute pancreatitis. Well-designed clinical and experimental research should be done. With the gradual elucidation of the regulatory mechanism of ICAM-1 and the pathogenic mechanism of acute lung injury, together with in-depth pharmacological research, and the diversification of treatment means, it is believed that the incidence rate of the disease will be reduced and the prognosis of acute pancreatitis patients will be significantly improved.

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References

1. Hartwig W, Werner J, Müller CA, Uhl W, Büchler MW. Surgical management of severe pancreatitis including sterile necrosis. *J Hepatobiliary Pancreat Surg* 2002; 9:429-35. [PMID 12483264]
2. Bai Y, Liu Y, Jia L, Jiang H, Ji M, Lv N, et al. Severe acute pancreatitis in China: etiology and mortality in 1976 patients. *Pancreas* 2007; 35:232-7. [PMID 17895843]
3. Surbatović M, Jovanović K, Radaković S, Filipović N. Pathophysiological aspects of severe acute pancreatitis-associated lung injury. *Srp Arh Celok Lek* 2005; 133:76-81. [PMID 16053182]
4. Liu XM, Xu J, Wang ZF. Pathogenesis of acute lung injury in rats with severe acute pancreatitis. *Hepatobiliary Pancreat Dis Int* 2005; 4:614-7. [PMID 16286275]
5. Bhatia M, Moochhala S. Role of inflammatory mediators in the pathophysiology of acute respiratory distress syndrome. *J Pathol* 2004; 202:145-56. [PMID 14743496]
6. Steer ML. Relationship between pancreatitis and lung diseases. *Respir Physiol* 2001; 128:13-6. [PMID 11535257]
7. Bella J, Kolatkar PR, Marlor CW, Greve JM, Rossmann MG. The structure of the two amino-terminal domains of human intercellular adhesion molecule-1 suggests how it functions as a rhinovirus receptor. *Virus Res* 1999; 62:107-17. [PMID 10507321]
8. Sun W, Watanabe Y, Wang ZQ. Expression and significance of ICAM-1 and its counter receptors LFA-1 and Mac-1 in experimental acute pancreatitis of rats. *World J Gastroenterol* 2006; 12:5005-9. [PMID 16937496]
9. Albelda SM, Smith CW, Ward PA. Adhesion molecules and inflammatory injury. *FASEB J* 1994; 8:504-12. [PMID 8181668]
10. van de Stolpe A, van der Saag PT. Intercellular adhesion molecule-1. *J Mol Med* 1996; 74:13-33. [PMID 8834767]
11. Inoue S, Nakao A, Kishimoto W, Murakami H, Harada A, Nonami T, Takagi H. LFA-1 (CD11a/CD18) and ICAM-1 (CD54) antibodies attenuate superoxide anion release from polymorphonuclear leukocytes in rats with experimental acute pancreatitis. *Pancreas* 1996; 12:183-8. [PMID 8720667]
12. Goliás C, Tsoutsis E, Matziridis A, Makridis P, Batistatou A, Charalabopoulos K. Review. Leukocyte and endothelial cell adhesion molecules in inflammation focusing on inflammatory heart disease. *In Vivo* 2007; 21:757-69. [PMID 18019409]
13. Roland CL, Harken AH, Sarr MG, Barnett CC Jr. ICAM-1 expression determines malignant potential of cancer. *Surgery* 2007; 141:705-7. [PMID 17560245]
14. Hartwig W, Werner J, Warshaw AL, Antoniu B, Castillo CF, Gebhard MM, et al. Membrane-bound ICAM-1 is upregulated by trypsin and contributes to leukocyte migration in acute pancreatitis. *Am J Physiol Gastrointest Liver Physiol* 2004; 287:G1194-9. [PMID 15308472]
15. Inoue K, Takano H, Yanagisawa R, Sakurai M, Shimada A, Yoshino S, et al. Protective role of urinary trypsin inhibitor in acute lung injury induced by lipopolysaccharide. *Exp Biol Med (Maywood)* 2005; 230:281-7. [PMID 15792950]
16. Brown K, Park S, Kanno T, Franzoso G, Siebenlist U. Mutual regulation of the transcriptional activator NF-kappa B and its inhibitor, I kappa B-alpha. *Proc Natl Acad Sci U S A* 1993; 90:2532-6. [PMID 8460169]
17. Baldwin AS Jr. The NF-kappa B and I kappa B proteins: new discoveries and insights. *Annu Rev Immunol* 1996; 14:649-83. [PMID 8717528]
18. Zhou Z, Connell MC, MacEwan DJ. TNFR1-induced NF-kappaB, but not ERK, p38MAPK or JNK activation, mediates TNF-induced ICAM-1 and VCAM-1 expression on endothelial cells. *Cell Signal* 2007; 19:1238-48. [PMID 17292586]
19. Norman J. The role of cytokines in the pathogenesis of acute pancreatitis. *Am J Surg* 1998; 175:76-83. [PMID 9445247]
20. Malleo G, Mazzone E, Siriwardena AK, Cuzzocrea S. TNF-alpha as a therapeutic target in acute pancreatitis--lessons from experimental models. *ScientificWorldJournal* 2007; 7:431-48. [PMID 17450307]
21. Zhang XP, Zhang L, Yang P, Zhang RP, Cheng QH. Protective effects of baicalin and octreotide on multiple organ injury in severe acute pancreatitis. *Dig Dis Sci* 2008; 53:581-91. [PMID 17549629]
22. van de Stolpe A, Caldenhoven E, Stade BG, Koenderman L, Raaijmakers JA, Johnson JP, van der Saag PT. 12-O-tetradecanoylphorbol-13-acetate- and tumor necrosis factor alpha-mediated induction of intercellular adhesion molecule-1 is inhibited by dexamethasone. Functional analysis of the human intercellular adhesion molecular-1 promoter. *J Biol Chem* 1994; 269:6185-92. [PMID 7907090]
23. Min JK, Kim YM, Kim SW, Kwon MC, Kong YY, Hwang IK, et al. TNF-related activation-induced cytokine enhances leukocyte adhesiveness: induction of ICAM-1 and VCAM-1 via TNF receptor-associated factor and protein kinase C-dependent NF-kappaB activation in endothelial cells. *J Immunol* 2005; 175:531-40. [PMID 15972689]
24. Masamune A, Shimosegawa T. Anti-cytokine therapy for severe acute pancreatitis. *Nippon Rinsho* 2004; 62:2116-21. [PMID 15552897]
25. Fujita M, Masamune A, Satoh A, Sakai Y, Satoh K, Shimosegawa T. Ascites of rat experimental model of severe acute pancreatitis induces lung injury. *Pancreas* 2001; 22:409-18. [PMID 11345143]
26. Martín Alonso MA, Santamaría A, Saracíbar E, Arranz E, Garrote JA, Almaraz A, Caro-Patón A. Cytokines and other immunological parameters as markers of distant organ involvement in acute pancreatitis. *Med Clin (Barc)* 2007; 128:401-6. [PMID 17394854]
27. Seely AJ, Naud JF, Campisi G, Giannias B, Liu S, DiCarlo A, et al. Alteration of chemoattractant receptor expression regulates human neutrophil chemotaxis in vivo. *Ann Surg* 2002; 235:550-9. [PMID 11923612]
28. Li YH, Huang ZW, Xue P, Guo J, He FQ, You Z, Wang ZR. Effects of Chaiqin Chengqi Decoction on activation of nuclear factor-kappa B in pancreas of rats with acute necrotizing pancreatitis. *Zhong Xi Yi Jie He Xue Bao* 2008; 6:180-4. [PMID 18241655]
29. Masamune A, Shimosegawa T, Fujita M, Satoh A, Koizumi M, Toyota T. Ascites of severe acute pancreatitis in rats transcriptionally up-regulates expression of interleukin-6 and -8 in vascular endothelium and mononuclear leukocytes. *Dig Dis Sci* 2000; 45:429-37. [PMID 10711463]
30. Takahashi HK, Iwagaki H, Hamano R, Kanke T, Liu K, Sadamori H, et al. Effect of adenosine receptor subtypes stimulation on mixed lymphocyte reaction. *Eur J Pharmacol* 2007; 564:204-10. [PMID 17374532]
31. Pereiaslov AA, Chuklin SM, Posivnych MM. Pathogenesis and treatment of hemocoagulation disorders in an acute necrotic pancreatitis. *Klin Khir* 2006; 10:26-9. [PMID 17269403]
32. Katakami N, Kaneto H, Matsuhisa M, Yoshiuchi K, Kato K, Yamamoto K, et al. Serum interleukin-18 levels are increased and closely associated with various soluble adhesion molecule levels in type 1 diabetic patients. *Diabetes Care* 2007; 30:159-61. [PMID 17192354]
33. Gloor B, Todd KE, Lane JS, Rigberg DA, Reber HA. Mechanism of increased lung injury after acute pancreatitis in IL-10 knockout mice. *J Surg Res* 1998; 80:110-4. [PMID 9790823]
34. Wang D, Jin D, Wu Z, Zou W, Xu D, Zheng Z, Liu X. Therapeutic effects of human interleukin 10 gene transfer on severe acute pancreatitis in rats, an experimental study. *Zhonghua Yi Xue Za Zhi* 2002; 82:844-7. [PMID 12126536]
35. Duran A, Diaz-Meco MT, Moscat J. Essential role of RelA Ser311 phosphorylation by zetaPKC in NF-kappaB transcriptional activation. *EMBO J* 2003; 22:3910-8. [PMID 12881425]
36. Hawkins BJ, Solt LA, Chowdhury I, Kazi AS, Abid MR, Aird WC, et al. G protein-coupled receptor Ca2+-linked mitochondrial

reactive oxygen species are essential for endothelial/leukocyte adherence. *Mol Cell Biol* 2007; 27:7582-93. [PMID 17724077]

37. Zhang XP, Wu CJ, Li ZJ. Advances in research of severe acute pancreatitis complicated by lung injury. *World Chinese Journal of Digestology* 2008; 16:299-306.

38. Melder RJ, Koenig GC, Witwer BP, Safabakhsh N, Munn LL, Jain RK. During angiogenesis, vascular endothelial growth factor and basic fibroblast growth factor regulate natural killer cell adhesion to tumor endothelium. *Nat Med* 1996; 2:992-7. [PMID 8782456]

39. Slogoff MI, Ethridge RT, Rajaraman S, Evers BM. COX-2 inhibition results in alterations in nuclear factor (NF)-kappaB activation but not cytokine production in acute pancreatitis. *J Gastrointest Surg* 2004; 8:511-9. [PMID 15120378]

40. Ten Kate M, Aalbers AG, Sluiter W, Hofland LJ, Sonneveld P, Jeekel J, Van Eijck CH. Polymorphonuclear leukocytes increase the adhesion of circulating tumor cells to microvascular endothelium. *Anticancer Res* 2007;27:17-22. [PMID 17352210]

41. Denham W, Yang J, Wang H, Botchkina G, Tracey KJ, Norman J. Inhibition of p38 mitogen activate kinase attenuates the severity of pancreatitis-induced adult respiratory distress syndrome. *Crit Care Med* 2000; 28:2567-72. [PMID 10921596]

42. Bhatnagar A, Wig JD, Majumdar S. Expression of activation, adhesion molecules and intracellular cytokines in acute pancreatitis. *Immunol Lett* 2001; 77:133-41. [PMID 11410245]

43. Hayashi T, Ishida Y, Kimura A, Iwakura Y, Mukaida N, Kondo T. IFN-gamma protects cerulein-induced acute pancreatitis by repressing NF-kappa B activation. *J Immunol* 2007; 178:7385-94. [PMID 17513789]

44. Kyriakides C, Jasleen J, Wang Y, Moore FD Jr, Ashley SW, Hechtman HB. Neutrophils, not complement, mediate the mortality of experimental hemorrhagic pancreatitis. *Pancreas* 2001; 22:40-6. [PMID 11138969]

45. Artigas A, Bernard GR, Carlet J, Dreyfuss D, Gattinoni L, Hudson L, et al. The American-European Consensus Conference on ARDS, part 2. Ventilatory, pharmacologic, supportive therapy, study design strategies and issues related to recovery and remodeling. *Intensive Care Med* 1998; 24:378-98. [PMID 9609420]

46. Aldridge AJ. Role of the neutrophil in septic shock and the adult respiratory distress syndrome. *Eur J Surg* 2002; 168:204-14. [PMID 12440757]

47. Sandoval D, Gukovskaya A, Reavey P, Gukovsky S, Sisk A, Braquet P, et al. The role of neutrophils and platelet-activating factor in mediating experimental pancreatitis. *Gastroenterology* 1996; 111:1081-91. [PMID 8831604]

48. Zhao X, Dib M, Wang X, Widegren B, Andersson R. Influence of mast cells on the expression of adhesion molecules on circulating and migrating leukocytes in acute pancreatitis-associated lung injury. *Lung* 2005; 183:253-64. [PMID 16211461]

49. Zhou MY, Lo SK, Bergenfeldt M, Tirupathi C, Jaffe A, Xu N, Malik AB. In vivo expression of neutrophil inhibitory factor via gene transfer prevents lipopolysaccharide-induced lung neutrophil infiltration and injury by a beta2 integrin-dependent mechanism. *J Clin Invest* 1998; 101:2427-37. [PMID 9616214]

50. Frossard JL, Saluja A, Bhagat L, Lee HS, Bhatia M, Hofbauer B, Steer ML. The role of intercellular adhesion molecule 1 and neutrophils in acute pancreatitis and pancreatitis-associated lung injury. *Gastroenterology* 1999; 116:694-701. [PMID 10029629]

51. Lu H, Ballantyne C, Smith CW. LFA-1 (CD11a/CD18) triggers hydrogen peroxide production by canine neutrophils. *J Leukoc Biol* 2000; 68:73-80. [PMID 10914492]

52. Folch E, Salas A, Panés J, Gelpí E, Roselló-Catafau J, Anderson DC, et al. Role of P-selectin and ICAM-1 in pancreatitis-induced lung inflammation in rats: significance of oxidative stress. *Ann Surg* 1999; 230:792-8. [PMID 10615934]

53. Walzog B, Weinmann P, Jeblonski F, Scharffetter-Kochanek K, Bommert K, Gaeltgens P. A role for beta(2) integrins (CD11/CD18) in the regulation of cytokine gene expression of polymorphonuclear

neutrophils during the inflammatory response. *FASEB J* 1999; 13:1855-65. [PMID 10506590]

54. Gao HK, Zhou ZG, Chen YD. Expression of platelet endothelial cell adhesion molecules-1 in leukocytes in pancreatic microcirculation of acute pancreatitis. *Chinese Journal of Experimental Surgery* 2002; 19:557-8.

55. Liu XM, Liu QG, Xu J, Pan CE. Microcirculation disturbance affects rats with acute severe pancreatitis following lung injury. *World J Gastroenterol* 2005; 11:6208-11. [PMID 16273652]

56. Raghu MG, Wig JD, Kochhar R, Gupta D, Gupta R, Yadav TD, et al. Lung complications in acute pancreatitis. *JOP. J Pancreas (Online)* 2007; 8:177-85. [PMID 17356240]

57. Folch-Puy E. Markers of severity in acute pancreatitis. *Med Clin (Barc)* 2007; 128:417-8. [PMID 17394857]

58. Kaufmann P, Smolle KH, Brunner GA, Demel U, Tilz GP, Krejs GJ. Relation of serial measurements of plasma-soluble intercellular adhesion molecule-1 to severity of acute pancreatitis. *Am J Gastroenterol* 1999; 94:2412-6. [PMID 10484001]

59. Foitzik T, Eibl G, Buhr HJ. Therapy for microcirculatory disorders in severe acute pancreatitis: comparison of delayed therapy with ICAM-1 antibodies and a specific endothelin A receptor antagonist. *J Gastrointest Surg* 2000; 4:240-6. [PMID 10769086]

60. Lundberg AH, Fukatsu K, Gaber L, Callicutt S, Kotb M, Wilcox H, et al. Blocking pulmonary ICAM-1 expression ameliorates lung injury in established diet-induced pancreatitis. *Ann Surg* 2001; 233:213-20. [PMID 11176127]

61. Yokomura I, Iwasaki Y, Nagata K, Nakanishi M, Natsuhara A, Harada H, et al. Role of intercellular adhesion molecule 1 in acute lung injury induced by candidemia. *Exp Lung Res* 2001; 27:417-31. [PMID 11480583]

62. Werner J, Hartwig W, Schmidt E, Gebhard MM, Herfarth C, Klar E. Reduction of local and systemic complications of acute pancreatitis by monoclonal antibody to ICAM-1. *Langenbecks Arch Chir Suppl Kongressbd* 1998; 115(Suppl 1):725-9. [PMID 14518349]

63. Rau B, Bauer A, Wang A, Gansauge F, Weidenbach H, Nevalainen T, et al. Modulation of endogenous nitric oxide synthase in experimental acute pancreatitis: role of anti-ICAM-1 and oxygen free radical scavengers. *Ann Surg* 2001; 233:195-203. [PMID 11176125]

64. Long J, Song N, Liu XP, Guo KJ, Guo RX. Nuclear factor-kappa B activation on the reactive oxygen species in acute necrotizing pancreatic rats. *World J Gastroenterol* 2005; 11:4277-80. [PMID 16015706]

65. Długosz JW, Andrzejewska A, Nowak K, Wróblewski E, Dabrowski A. The cumulative effect of nuclear factor-kappaB (NF-kappaB) inhibition and endothelins in early cerulein-induced acute pancreatitis in rats. *Rocz Akad Med Białymst* 2005; 50:230-6. [PMID 16358973]

66. Shi C, Zhao X, Lagergren A, Sigvardsson M, Wang X, Andersson R. Immune status and inflammatory response differ locally and systemically in severe acute pancreatitis. *Scand J Gastroenterol* 2006; 41:472-80. [PMID 16635917]

67. Virlos I, Mazzon E, Serraino I, Genovese T, Di Paola R, Thiemerman C, et al. Calpain I inhibitor ameliorates the indices of disease severity in a murine model of cerulein-induced acute pancreatitis. *Intensive Care Med* 2004; 30:1645-51. [PMID 15168010]

68. Virlos I, Mazzon E, Serraino I, Di Paola R, Genovese T, Britti D, et al. Pyrrolidine dithiocarbamate reduces the severity of cerulein-induced murine acute pancreatitis. *Shock* 2003; 20:544-50. [PMID 14625479]

69. Satoh A, Shimosegawa T, Fujita M, Kimura K, Masamune A, Koizumi M, Toyota T. Inhibition of nuclear factor-kappaB activation improves the survival of rats with taurocholate pancreatitis. *Gut* 1999; 44:253-8. [PMID 9895386]

70. Jaffray C, Yang J, Carter G, Mendez C, Norman J. Pancreatic elastase activates pulmonary nuclear factor kappa B and inhibitory

kappa B, mimicking pancreatitis-associated adult respiratory distress syndrome. *Surgery* 2000; 128:225-31. [PMID 10922996]

71. Hietaranta AJ, Saluja AK, Bhagat L, Singh VP, Song AM, Steer ML. Relationship between NF-kappaB and trypsinogen activation in rat pancreas after supramaximal caerulein stimulation. *Biochem Biophys Res Commun* 2001; 280:388-95. [PMID 11162528]

72. Li SL, Chen X, Wu T, Liu JD. Protective effect of proteasome inhibitor MG-132 in rats with severe acute pancreatitis and lung injury. *Nan Fang Yi Ke Da Xue Xue Bao* 2007; 27:1845-7. [PMID 18158999]

73. Letoha T, Fehér LZ, Pecze L, Somlai C, Varga I, Kaszaki J, et al. Therapeutic proteasome inhibition in experimental acute pancreatitis. *World J Gastroenterol.* 2007;13:4452-7. [PMID 17724800]

74. Chen X, Li SL, Wu T, Liu JD. Proteasome inhibitor ameliorates severe acute pancreatitis and associated lung injury of rats. *World J Gastroenterol* 2008; 14:3249-53. [PMID 18506934]

75. Shimizu K, Shiratori K, Hayashi N, Kobayashi M, Fujiwara T, Horikoshi H. Thiazolidinedione derivatives as novel therapeutic agents to prevent the development of chronic pancreatitis. *Pancreas* 2002; 24:184-90. [PMID 11854624]

76. Xu P, Zhou XJ, Chen LQ, Chen J, Xie Y, Lv LH, Hou XH. Pioglitazone attenuates the severity of sodium taurocholate-induced severe acute pancreatitis. *World J Gastroenterol* 2007, 13:1983-8. [PMID 17461502]

77. Cuzzocrea S, Pisano B, Dugo L, Ianaro A, Britti D, Patel NS, et al. Rosiglitazone, a ligand of the peroxisome proliferator-activated receptor-gamma, reduces acute pancreatitis induced by cerulein. *Intensive Care Med* 2004; 30:951-6. [PMID 14985957]

78. Ivashchenko CY, Duan SZ, Usher MG, Mortensen RM. PPAR-gamma knockout in pancreatic epithelial cells abolishes the inhibitory effect of rosiglitazone on caerulein-induced acute pancreatitis. *Am J Physiol Gastrointest Liver Physiol* 2007; 293:G319-26. [PMID 17463185]

79. Pasceri V, Wu HD, Willerson JT, Yeh ET. Modulation of vascular inflammation in vitro and in vivo by peroxisome proliferator-activated receptor-gamma activators. *Circulation* 2000; 101:235-8. [PMID 10645917]

80. Wang P, Anderson PO, Chen S, Paulsson KM, Sjögren HO, Li S. Inhibition of the transcription factors AP-1 and NF-kappaB in CD4 T cells by peroxisome proliferator-activated receptor gamma ligands. *Int Immunopharmacol* 2001; 1:803-12. [PMID 11357893]

81. Ricote M, Li AC, Willson TM, Kelly CJ, Glass CK. The peroxisome proliferator-activated receptor-gamma is a negative regulator of macrophage activation. *Nature* 1998; 391:79-82. [PMID 9422508]

82. Quinlan KL, Song IS, Bunnett NW, Letran E, Steinhoff M, Harten B, et al. Neuropeptide regulation of human dermal microvascular endothelial cell ICAM-1 expression and function. *Am J Physiol* 1998, 275:C1580-90. [PMID 9843720]

83. Lau HY, Bhatia M. Effect of CP-96,345 on the expression of adhesion molecules in acute pancreatitis in mice. *Am J Physiol Gastrointest Liver Physiol* 2007; 292:G1283-92. [PMID 17218475]

84. Reed KL, Fruin AB, Gower AC, Gonzales KD, Stucchi AF, Andry CD, et al. NF-kappaB activation precedes increases in mRNA encoding neurokinin-1 receptor, proinflammatory cytokines, and adhesion molecules in dextran sulfate sodium-induced colitis in rats. *Dig Dis Sci* 2005; 50:2366-78. [PMID 16416193]