HIGHLIGHT ARTICLE

Advances in Immunotherapy for Pancreatic Cancer: 2013 Highlights from the "2013 ASCO Annual Meeting". Chicago, IL, USA; May 30 - June 4, 2013

Nicholas C DeVito, Muhammad Wasif Saif

Tufts Medical Center. Boston, MA, USA

Summary

Pancreatic cancer is one of the more difficult malignancies to treat, and there is a great need for less toxic, effective regimens. Immunotherapy has shown potential in the treatment of pancreatic cancer, and at ASCO 2013 there were several progressive advances in its clinical application. Abstracts #3067, #3049, #3007, #4040, #LBA4004, and #3090 will be discussed. New developments in the field of immunotherapy are promising novel treatments for pancreatic neoplasms with tolerable side effect profiles.

What Did We Know Before the 2013 American Society of Clinical Oncology (ASCO) Annual Meeting?

Pancreatic cancer remains the fourth leading cause of cancer related deaths in the USA, with 45,220 estimated new cases in 2013 and 38,460 deaths [1]. The 5-year survival remains less than 5% for patients with metastatic pancreatic adenocarcinoma [2]. Unfortunately, the majority of pancreatic cancers are diagnosed when the disease is already metastatic or locally invasive. Metastatic pancreatic cancer has a survival of less than 20% at year. one While surgical resection and chemotherapy improve survival in early stage disease, post-resection survival at 5 years remains 15-20% [3]. The few chemotherapeutic agents that have shown efficacy in the treatment of pancreatic cancer include gemcitabine and FOLFIRINOX (a combination of 5-fluorouracil. leucovorin. oxaliplatin, and irinotecan) [4, 5]. Chemoradiation has a modest benefit in unresectable disease that is locally advanced. However, despite many advances the survival for patients with pancreatic cancer remains poor. Frequently, diagnosis is made at a

Key wordsPancreatic Neoplasms, immunotherapy, vaccine,
chemoimmunotherapyAbbreviationsCY:
cyclophosphamide;FOLFIRINOX:5-

fluorouracil, leucovorin, oxaliplatin, and irinotecan; HSP: heat shock protein; LAK: lymphokine activated killer; MSLN: membrane-bound recombinant mesothelin; rAd-p53: recombinant adenovirus-p53 **Correspondence** Nicholas C DeVito Tufts Medical Center; 800 W. Washington St; Boston, MA 0211;

Tufts Medical Center; 800 W. Washington St; Boston, MA 0211 USA Phone: +1-617.636.5000; Fax: +1-617.636.8538 E-mail: ndevito@tuftsmedicalcenter.org late stage, which makes interventions difficult; therefore, we are in dire need of better options for these patients.

Immunotherapy offers a novel approach that has the potential to function alone or in concert with traditional therapies. Recently, multiple modalities have been shown to improve survival without adding toxicity. Examples of the different vaccines that have been in development for pancreatic cancer are demonstrated in Table 1, the most advanced of which being algenpantucel-L.

In addition to the aforementioned methods, there has been an interest in antigen-pulsed dendritic cells. Dendritic cells are the most capable antigen presenting cell of priming naive T cells, stimulating memory T cells to upregulate an antigen specific response. When dendritic cell based immunotherapy with gemcitabine was given to 49 patients with late stage pancreatic adenocarcinoma in a phase I trial, with or without lymphokine activated killer (LAK) therapy, patients with receiving LAK along with chemotherapy and antigen pulsed dendritic cells had prolonged survival. This therapy was also deemed to be safe [32]. Antigen targets like CEA [33] and mucin 1 [34, 35, 36] are safe by phase I investigation, but have not gone beyond this stage. Lastly, heat shock protein (HSP), an intracellular peptide chaperone induced by stress has also been a target of immunotherapy. HSPPC-96 is an autologous HSPbased vaccine produced from resected tumor tissue, and has been shown to induce immune response [37]. A phase I study determined its safety [38]. Both of these methods require further studies to determine their clinical efficacy.

What Have We Learnt from the 2013 American Society of Clinical Oncology (ASCO) Annual Meeting?

At this year's ASCO Annual Meeting, we can see that there is an exponentially expanding interest in the field of immunotherapy, especially directed at pancreatic adenocarcinoma. A basic overview of the abstracts presented is listed in Table 2. Particular abstracts will be examined in detail, with future directions as well as current clinical trials left for the discussion section.

<u>A Phase III Randomized Trial of Chemoimmunotherapy Comprising Gemcitabine and Capecitabine</u> with or without Telomerase Vaccine GV1001 in Patients with Locally Advanced or Metastatic Pancreatic Cancer (Abstract #LBA4004 [26])

The concept of adding the telomerase vaccine GV1001, targeting a class II (T helper) epitope expressed in 90% of pancreatic cancers, with gemcitabine, was based off the perception that gemcitabine has been shown in pre-clinical studies to increase antigen cross-presentation, enhance T cell activation and tumor-infiltrating lymphocytes

as well as reduce myeloid-derived suppressor cells and T regulatory cells [44, 45, 46, 47]. Onethousand and 110 patients with advanced pancreatic cancer were randomized to three arms: Arm 1: receiving gemcitabine and capecitabine (GemCap); Arm 2: receiving GemCap followed by GV1001 and additional GemCap; and Arm 3: concurrent GemCap and GV1001. Response rates were 17.6%, 8.9% and 15.5% in Arm 1, Arm 2, and Arm 3, respectively. It was then concluded that overall survival with concurrent GemCap and GV1001 was not improved over GemCap alone, and sequential administration of GV1001 with GemCap before and after (if tolerated by patients) did not demonstrate significantly improved survival either. The addition of GV1001 to GemCap did not improve outcome of the patients in this large clinical trial.

Interim Safety ar	<u>ıd Efficac</u>	<u>y Anal</u>	lysis of	<u>a Phase II</u>
Randomized Stud	ly of GVA	X Pand	creas a	nd CRS-207
Immunotherapy	in Pat	ients	with	Metastatic
Pancreatic Cancer	· (Abstrac	t #404(<u>0 [43])</u>	

CRS-207 is a live-attenuated *Listeria monocytogenes* engineered to express human mesothelin shown to be safe in a phase I trial.

Table 1. Current vaccines for pancreatic cancer utilized in human studies.

Class of vaccine	Description	Summary of trials	Outcome	Future advances
Whole-cell vaccines Combine dendritic cells with antigen to present to patient leukocytes	Algenpantucel-L: Irradiated, live combination of two human allogeneic pancreatic cancer cells that express murine alpha-1,3-galactosyl transferase which are absent in humans. Antibodies are expressed leading to complement activation and cell mediated immunity directed at tumor antigens [6, 7, 8, 9, 10]	Hardacre 2012, Phase II 73 patients receiving 100 or 300 million cells in subdermal injections with adjuvant gemcitabine and 5-FU post R0/1 resection [11]	Observed survival of 86% compared to predicted 1 year of 55- 63%	Phase III multicenter trial
	GM-CSF vaccine (GVAX): Irradiated tumor cells expressing GM-CSF first described in melenoma [12]	Jaffee 2001, 14 patients with stage I-III PanCa post resection with chemoradiation [13]	Improved outcome in delayed type hyper- sensitivity responders	Trials in patients with more advance staged pancreatic
		Lutz 2011, phase II, 60 patients post resection [14]	1-year survival: 85%	cancer that is unresectable, metastatic
Peptide and DNA vaccines	Ras peptide vaccine: Common cancer mutation, targetable by CD4/8 T cells [15]. Peptide vaccine [16]	Phase I trials: Gjertsen 1996, 5 patients unresectable [17]; Carbone 2005, 39 patients [18]	Safe; some patients had immune response	N/A
		Gjertsen 2001, Phase I/II, 48 patients [19]	20 patients with response, 4 survived vs. 0 of non-vaccinated group	
	Telomerase peptide vaccine (GV1001): Expressed in 90% of cancer cells [20]. Telomerase maintains	Bernhardt 2006, Phase I/II, 48 patients given GV1001 + GM-CSF [25]	Tolerated, immune response correlated with survival	TeloVac Phase III trial
	chromosome stability; mutations render cells immortal. Expressed in PanCa [21, 22, 23, 24]	Phase III, Primo Vac vaccine monotherapy <i>vs</i> . gemcitabine [26]	Closed early due to lack of survival advantage	
	CEA and Mucin 1 (TRICOM): Poxvirus based vaccine containing B7-1, ICAM-1 and LFA-3 [27]	Phase I studies [27]	Safe, significant survival benefit in patients with immune response	N/A
		Phase III PANVAC-V/F [28, 29]	Did not improve survival	
	Survivin: apoptosis inhibitor, upregulated in PanCa [30]	Case report [31]	Complete response in one patient; disease recurrence when vaccine stopped	N/A

GM-CSF: granulocyte-macrophage colony stimulating factor; ICAM-1: intercellular adhesion molecule; LFA-3: leukocyte function associated antigen 3; N/A: not applicable; PanCa: pancreatic cancer

Table 2	. Abstracts	presente	d at the	e 2013 ASC	0 Annula Meeting	relating to v	accine thera	py for pancreatic can	cer
	-		-			_		-	

Author; abstract	Study design	Country	Patients	Treatment	Outcome	Grade 3/4 adverse events
Schmitz-Winnenthal, et al.; #3090 [39]	Phase I	Germany	45 patients, advance PanCa	VXM01 <i>vs</i> . placebo	Safe, induces and enhances VEGFR-2 T cell response	Leukopenia, abdominal pain, diarrhea (also in placebo)
Bae <i>et al.</i> #3067 [40]	Cellular study	USA	HLA-A2 breast, colon, pancreatic cell lines	XBP1	Invoke effector memory responses	N/A
Chai, <i>et al.</i> #3049 [41]	Phase II	China	36 Stage IV PanCa	Dendritic cell ± recombinant adenovirus p53	Combination beneficial by partial response and stable disease, safe	None
Rossi, <i>et al.</i> #3007 [42] (multicenter)	Phase II	USA	64 patients with resected PanCa cancer	Algenpantucel-L	Anti-MSLN antibody correlates with improved overall survival	Not indicated in abstract
Le, <i>et al</i> . #4040 [43]	Phase II	USA	90 patients, metastatic PanCa, median 3 prior regimens	CY/GVAX ± CRS- 207	Significant survival difference. Well tolerated	Fever, lymphopenia, hypophasphatemia
Middleton, <i>et al.</i> #LBA4004 [26]		I England	1,062 patients	GemCap ± GV1001	No overall survival difference when GV1001 added	Not reported

GemCap: gemcitabine/capecitabine; N/A not applicable; PanCa: pancreatic cancer

Additionally, using both GVAX and CRS-207 has shown synergy in mouse models [48, 49]. Prior studies showed that GVAX with cyclophosphamide (CY/GVAX) induced mesothelin-specific T cell responses improved survival in patients with metastatic pancreatic cancer [14]. This study also cites a phase I trial that demonstrated anecdotal survival difference in CRS-207 with patients who had received GVAX [49]. Ninety patients were treated with either: Arm A: 2 doses of CY/GVAX followed by 4 doses of CRS-207; or Arm B: 6 doses of CY/GVAX every 3 weeks. No serious adverse events were observed, though grade 3/4 fever, lymphopenia, hypophosphatemia, elevated liver enzymes and fatigue were seen in less than 5% of subjects receiving CRS-207. Fifty-one patients of the original 90 were evaluated post-treatment; 34% had stable disease in Arm A compared to 19% in Arm B. Overall survival was 6 months in Arm A while 3.4 months in Arm B. This lead to the conclusion that CY/GVAX with CRS-207 was not only well tolerated, but the observed overall survival difference was significant enough to lead to early stopping of the trial.

<u>Phase II Study of Dendritic Cell Vaccination Combined</u> <u>with Recombinant Adenovirus p53 in Treatment for</u> <u>Patients with Advanced Pancreatic Carcinoma</u> <u>(Abstract #3049 [41])</u>

This study compared both the clinical response and immunological response of 36 patients with stage IV pancreatic cancer receiving dendritic cell vaccines with or without recombinant adenovirusp53 (rAd-p53). The investigators measured the expression of CD31, CD41, and CD41/CD81 ratio in peripheral blood, which all increased in both groups, while a decrease in T regulatory cells was observed. Clinical response was significant, as patients receiving rAd-p53 had a disease control rate of 45% (compared to 37.5% in those not receiving rAd-p53), a 6-month overall survival of 50% (compared to 43.8%) and median survival of 6.8 months as opposed to 5.5 months. Only mild to medium grade fever was observed, and no serious adverse events occurred. This study concluded that dendritic cell-based immunotherapy was safe, and that the combination with rAd-p53 was beneficial to survival and disease control.

Effect of Algenpantucel-L Immunotherapy for Pancreatic Cancer on Anti-Mesothelin Antibody (Ab) Titers and Correlation with Improved Overall Survival (Abstract #3007 [42])

Using the algenpantucel-L in a multicenter phase II study with gemcitabine, 5-fluorouracil and chemoradiotherapy (the standard of care) in 64 patients with resected pancreatic cancer, the authors measured the clinical response as well as IgG, complement, CA 19-9, anti-aGal antibody, anti-CEA antibody and most importantly, antimembrane-bound recombinant mesothelin (MSLN) antibody. Disease free survival was 62% with a 12month overall survival of 86% in the setting of a 3year follow up. Overall survival at 3 years was 39% while disease free survival was 26%. Thirty-one percent of patients had increased anti-MSLN antibody and notably had a median overall survival of 42 months compared to 20 months in those who did not seroconvert.

Discussion

Pancreatic cancer is an exceedingly difficult disease to treat, with recent advances only showing a modest survival benefit. There is a great need for novel therapies, preferably those with favorable side effect profiles. Immunotherapy is an everexpanding field offering a multitude of targeted agents that show promise in the treatment of solid tumors, including pancreatic neoplasms. There is yet to be an FDA approved vaccine for pancreatic cancer. We have seen some major progressions in the field of cancer immunotherapy, as exemplified in the abstracts presented at the 2013 ASCO Annual Meeting.

Overall survival is the ultimate outcome measure for immunotherapy in pancreatic cancer, especially given the dismal prognosis in advance diseases. However, it is important to assess immune response to correlate to clinical benefit, as seen in the Rossi study presented as ASCO regarding anti-MSLN antibody seroconversion in patients given the algenpantucel-L vaccine. The identification of these targets is critical to developing new therapies and monitoring immune activation. Pancreatic cancer antigens that have been identified include aGal epitopes, mesothelin, VEGFR, telomerase and the addition of XBP1 at ASCO2013.

The immune response to cancer in general is clearly blunted by the downregulation of T, B and NK cell response from T regulatory cells and MDSCs. A recurring theme in these abstracts is the addition of chemotherapy with vaccines, which although seemingly counterintuitive as chemotherapy is generally thought of as immunosuppressant; this is not the case. Multiple chemotherapeutic agents have been shown to expose intracellular tumor antigens resulting in improved antigen-specific CD8+ T cell response, decrease T regulatory cells and MDSCs, and may enhance Th1 response. Eliminating T-regs and MDSCs can bring about a more robust, effective tumor-specific immune reaction, as seen in trials using gemcitabine (Figure 1) [50]. This leads to the conclusion that there is a great benefit to chemotherapy in concert with immunotherapy (Figure 2) [44, 45, 46, 47].

Probably one of the most prominent findings from ASCO 2013 is a large Phase III trial using the

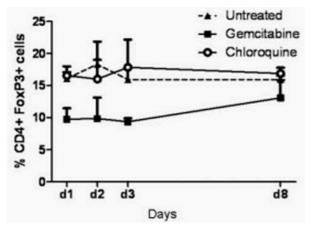


Figure 1. Quantification of mouse T regulatory cells after administration of gemcitabine. (Adapted from Rettig *et al.*, 2011 [50]. This figure is reproduced with permission of John Wiley & Sons, Inc.).

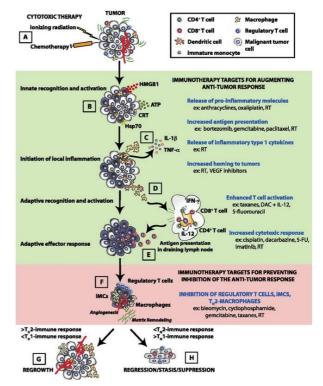


Figure 2. Immune response pathways induced following cytotoxic therapy. (Adapted from Shiao *et al.*, 2011 [47], with permission).

telomerase peptide vaccine GV1001 showing no survival advantage. Despite the theoretical applications of a vaccine targeting telomerase, it has not moved forward clinically; however there are still ongoing investigations and optimism [51].

The most promising and advanced therapies include GVAX and Algenpantucel-L. GVAX, when combined with cyclophosphamide is known to be a safe therapy with survival advantage. The addition of a modified *Listeria monocytogenes* with CRS-207 lead to no additional side effects but an improved survival difference when immunological response was achieved. Algenpantucel-L (currently in multicenter phase III trials) was shown to have enhanced effect when patients seroconverted and Anti-MSLN antibodies were detected; this could prove to be a valuable marker of vaccine efficacy in the future.

The addition of an attenuated adenovirus expressing p53 to a dendritic cell vaccine was also shown to enhance immune response and stabilize disease while also remaining a safe treatment. Viral vector trials are currently underway in the US and could offer an adjunct to immunotherapy, enhancing and broadening the immune response. Other therapies that are still in their infancy include an intriguing peptide vaccine which induces VEGFR-2 T-cell response. It was shown to be safe and should be moving into larger trials. There could be an interest in combining this regimen with

Table 3. Ongoing trials in vaccines for pancreatic cancer.	
---	--

Investigator/Center	Stage	Phase	Vaccine	Antigen
Chin/MUSC	Unresectable PanCa	Ι	Poly ICLC + dendritic cells	Autologous dendritic cells
Herman/Johns Hopkins	Resectable PanCa	Ι	CY/GVAX + FOLFIRINOX, SBRT	Irradiated Whole cell w/GM-CSF
Laheru/Johns Hopkins	Resectable, Stage I or II PanCa	Ι	GVAX ± CY	Irradiated Whole cell w/GM-CSF
Hausman/Oncothyreon	III or IV solid tumors	Ι	ONT-10	MUC1 Peptide vaccine
Schmitz-Winnenthal/ VAXIMM GmbH	Stage IV PanCa	Ι	VXM01	VEGFR-2 DNA vaccine
Chung/NCI	Stage IV Colon, Rectal, PanCa	Ι	MVAp53	p53 viral vector
Poplin/NCI	Unresectable PanCa	Ι	PANVAC-F, PANVAC-V with GM-CSF	CEA and MUC1 viral vectors
Becker/ Julius Maximilians Universitaet Hospital	Melanoma, PanCa, Colon, Cervical	I/II	Survivin peptide vaccine	Survivin
Zheng/Johns Hopkins	Resected PanCa	II	GVAX with boost vaccinations ± CY	Irradiated whole cell w/GM-CSF
Vahanian/NewLink (multicenter trial)	Resected PanCa	III	Standard of care (gemcitabine with or without 5-FU, chemoradiation) ± algenpantucel-L	Genetically modified aGal expressing PanCa cells

CY: cyclophosphamide; GM-CSF: granulocyte-macrophage colony stimulating factor; MVAp53: modified vaccinia virus Ankara expressing p53; PanCa: pancreatic cancer; SBRT: stereotactic body radiation therapy

chemotherapy like gemcitabine or cyclophosphamide, as this may in enhance immune response by the previously mentioned mechanisms.

In addition to chemotherapy, other ways to possibly enhance vaccines in the future could include comparing the use of different adjuvant, like toll-like receptor agonists [52], as well as the addition of immunomodulators like ipilimumab (anti-CTLA-4), nivolumab (Anti-PD1), lucatumumab (anti-CD40) and interleukins. Ipilimumab has been unsuccessful as a single agent [53] but is currently being examined with alganpantucel-L by Le *et al.* at Johns Hopkins (NCT00836407 – http://clinicaltrials. gov). Anti-CD40 boosts CD8+ T cell response in murine models of pancreatic cancer, but has not been utilized in human models [54]. These adjuncts to immunotherapy may help to overcome anergy in cytotoxic T lymphocytes and create a more vigorous immune response, which has been trialed in other cancers and yet to be examined in pancreatic adenocarcinoma [55, 56, 57]. Current ongoing trials can be seen in Table 3.

Conclusions

To conclude, we can see that there is a strong rise in immunotherapy options for resilient cancers like pancreatic adenocarcinoma, yet there are many hurdles to overcome. It is becoming clear that we must not only find antigens to target, but decrease the tumor's ability to downregulate the immune response. Once this is achieved, immune recognition by cytotoxic T cells can be greatly enhanced by targeted vaccines, leading to tumor apoptosis and optimally a clinically significant response. It is our hope that immunotherapy will provide a survival benefit without adding toxicity to chemotherapy regimens. Optimally, it will produce a long lasting immune memory response that can be correlated with longer term survival with the need for fewer cycles of chemotherapy, namely in patients who could not tolerate the side effects of traditional chemotherapy. There is still a great body of work in this field, but we have a myriad of safe options that could be advantageous to enhancing survival without increasing suffering in patients with this devastating disease.

Conflicts of interest The authors have no conflicts to disclose

References

1. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. CA Cancer J Clin. 2013;63(1):11-30. doi: 10.3322/caac.21166; 10.3322/caac.21166.

2. Jemal, A., Bray, F., Center, M. M., Ferlay, J., Ward, E. and Forman, D. Global cancer statistics. CA: A Cancer Journal for Clinicians, 2011; 61: 69–90.

3. Bilimoria KY, Bentrem DJ, Ko CY, et al. Validation of the 6th edition AJCC pancreatic cancer staging system: Report from the national cancer database. Cancer. 2007; 110(4):738-744. doi: 10.1002/cncr.22852.

4. Conroy, T., Desseigne, F., Ychou, M., Bouche, O., Guimbaud, R., Becouarn, Y. et al. FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. N Engl J Med 2011; 364: 1817–1825

5. Oettle, H., Post, S., Neuhaus, P., Gellert, K., Langrehr, J., Ridwelski, K. et al. Adjuvant chemotherapy with gemcitabine vs. observation in patients undergoing curative-intent resection of pancreatic cancer: a randomized controlled trial. JAMA 2007; 297: 267–277.

6. Galili U, Clark MR, Shohet SB, Buehler J, Macher BA. Evolutionary relationship between the natural anti-gal antibody and the gal alpha 1----3Gal epitope in primates. Proc Natl Acad Sci U S A. 1987; 84(5):1369-1373.

7. Galili U, Shohet SB, Kobrin E, Stults CL, Macher BA. Man, apes, and old world monkeys differ from other mammals in the expression of alpha-galactosyl epitopes on nucleated cells. J Biol Chem. 1988; 263(33):17755-17762.

8. Rossi, G., Mautino, M., Awwad, D., Husske, K., Lejukole, H., Koenigsfeld, M. et al. Allogeneic melanoma vaccine expressing alphaGal epitopes induces antitumor immunity to autologous antigens in mice without signs of toxicity. J Immunother 2008; 31: 545–554.

9. Rossi, G., Mautino, M., Unfer, R., Seregina, T., Vahanian, N. and Link, C. Effective treatment of preexisting melanoma with whole cell vaccines expressing alpha(1,3)-galactosyl epitopes. Cancer Res 2005; 65: 10555–10561.

10. Rossi, G., Unfer, R., Seregina, T. and Link, C. Complete protection against melanoma in absence of autoimmune depigmentation after rejection of melanoma cells expressing alpha(1,3)galactosyl epitopes. Cancer Immunol Immunother 2005; 54: 999–1009.

11. Hardacre, J., Mulcahy, M., Small, W., Talamonti, M., Obel, J. and Lima, R. Addition of algenpantucel-L immunotherapy to standard of care (SOC) adjuvant therapy for pancreatic cancer. J Clin Oncol 2012; 30(Suppl.): abstract 4049.

12. Dranoff, G., Jaffee, E., Lazenby, A., Golumbek, P., Levitsky, H., Brose, K. et al. Vaccination with irradiated tumor cells engineered to secrete murine granulocyte-macrophage colonystimulating factor stimulates potent, specific, and long-lasting anti-tumor immunity. Proc Natl Acad Sci U S A 1993; 90: 3539– 3543.

13. Jaffee, E., Hruban, R., Biedrzycki, B., Laheru, D., Schepers, K., Sauter, P. et al. Novel allogeneic granulocyte-macrophage colonystimulating factorsecreting tumor vaccine for pancreatic cancer: a phase I trial of safety and immune activation. J Clin Oncol 2001; 19: 145–156.

14. Lutz, E., Yeo, C., Lillemoe, K., Biedrzycki, B., Kobrin, B., Herman, J. et al. A lethally irradiated allogeneic granulocytemacrophage colony stimulating factor-secreting tumor vaccine for pancreatic adenocarcinoma. A phase II trial of safety, efficacy, and immune activation. Ann Surg 2011; 253: 328–335.

15. Abrams, S., Khleif, S., Bergmann-Leitner, E., Kantor, J., Chung, Y., Hamilton, J. et al. Generation of stable CD4+ and CD8+ T cell lines from patients immunized with ras oncogene-derived peptides reflecting codon 12 mutations. Cell Immunol 1997; 182: 137–151.

16. Gjertsen, M., Bakka, A., Breivik, J., Saeterdal, I., Solheim, B., Soreide, O. et al. Vaccination with mutant ras peptides and induction of T-cell responsiveness in pancreatic carcinoma patients carrying the corresponding RAS mutation. Lancet 1995; 346: 1399–1400.

17. Gjertsen, M., Bakka, A., Breivik, J., Saeterdal, I., Gedde-Dahl, T., Stokke, K. 3rd et al. Ex vivo ras peptide vaccination in patients with advanced pancreatic cancer: results of a phase I/II study. Int J Cancer 1996; 65: 450–453.

18. Carbone, D., Ciernik, I., Kelley, M., Smith, M., Nadaf, S., Kavanaugh, D. et al. Immunization with mutant p53- and K-rasderived peptides in cancer patients: immune response and clinical outcome. J Clin Oncol 2005; 23: 5099–5107.

19. Gjertsen, M., Buanes, T., Rosseland, A., Bakka, A., Gladhaug, I., Soreide, O. et al. Intradermal ras peptide vaccination with granulocyte–macrophage colony-stimulating factor as adjuvant: clinical and immunological responses in patients with pancreatic adenocarcinoma. Int J Cancer 2001; 92: 441–450.

20. Vasef, M., Ross, J. and Cohen, M. Telomerase activity in human solid tumors. Diagnostic utility and clinical applications. Am J Clin Pathol 1999; 112(1 Suppl. 1): S68–S75.

21. Hiyama E, Kodama T, Shinbara K, et al. Telomerase activity is detected in pancreatic cancer but not in benign tumors. Cancer Res. 1997; 57(2):326-331

22. Mizumoto K, Tanaka M. Detection of telomerase activity in patients with pancreatic cancer. Methods Mol Med. 2005; 103:199-205

23. Suehara, N., Mizumoto, K., Kusumoto, M., Niiyama, H., Ogawa, T., Yamaguchi, K. et al. Telomerase activity detected in pancreatic

juice 19 months before a tumor is detected in a patient with pancreatic cancer. Am J Gastroenterol 1998; 93: 1967–1971.

24. Kyte JA. Cancer vaccination with telomerase peptide GV1001. Expert Opin Investig Drugs. 2009;18(5):687-694.

25. Bernhardt, S., Gjertsen, M., Trachsel, S., Moller, M., Eriksen, J., Meo, M. et al. Telomerase peptide vaccination of patients with non-resectable pancreatic cancer: A dose escalating phase I/II study. Br J Cancer 2006; 95: 1474–1482.

26. Middleton GW, Valle JW, Wadsley J, Propper D, Coxon FY, Ross PJ, et al. A phase III randomized trial of chemoimmunotherapy comprising gemcitabine and capecitabine with or without telomerase vaccine GV1001 in patients with locally advanced or metastatic pancreatic cancer. J Clin Oncol 2103; 31(Suppl.): Abstract #LBA4004.

27. Kaufman, H., Kim-Schulze, S., Manson, K., DeRaffele, G., Mitcham, J., Seo, K. et al. Poxvirus-based vaccine therapy for patients with advanced pancreatic cancer. J Transl Med 2007; 5: 60.

28. Leuchtenberger M. Therion Biologics Corporation Reports Results of Phase 3 PANVAC-VF Trial and Announces Plans for Company Sale. BioSpace, Inc., Centennial, CO, USA;2006 (Accessed June 6th, 2013. <u>http://www.biospace.com/News/</u> therion-biologics-corporation-reports-results-of/23073

29. Madan RA, Arlen PM, Gulley JL. PANVAC-VF: poxviral-based vaccine therapy targeting CEA and MUC1 in carcinoma. Expert Opin Biol Ther. 2007 Apr;7(4):543-54.

30. Zhu, K., Qin, H., Cha, S., Neelapu, S., Overwijk, W., Lizee, G. et al. Survivin DNA vaccine generated specific antitumor effects in pancreatic carcinoma and lymphoma mouse models. Vaccine 2007; 25: 7955–7961.

31. Wobser, M., Keikavoussi, P., Kunzmann, V., Weininger, M., Andersen, M. and Becker, J. Complete remission of liver metastasis of pancreatic cancer under vaccination with a HLA-A2 restricted peptide derived from the universal tumor antigen survivin. Cancer Immunol Immunother 2006; 55: 1294–1298.

32. Kimura, Y., Tsukada, J., Tomoda, T., Takahashi, H., Imai, K., Shimamura, K. et al. Clinical and immunologic evaluation of dendritic cell-based immunotherapy in combination with gemcitabine and/ or S-1 in patients with advanced pancreatic carcinoma. Pancreas 2012; 41: 195–205.

33. Morse, M., Nair, S., Boczkowski, D., Tyler, D., Hurwitz, H., Proia, A. et al. The feasibility and safety of immunotherapy with dendritic cells loaded with CEA mRNA following neoadjuvant chemoradiotherapy and resection of pancreatic cancer. Int J Gastrointest Cancer 2002; 32: 1–6.

34. Pecher, G., Haring, A., Kaiser, L. and Thiel, E. Mucin gene (MUC1) transfected dendritic cells as vaccine: results of a phase I/II clinical trial. Cancer Immunol Immunother 2002; 51: 669–673.

35. Ramanathan, R., Lee, K., McKolanis, J., Hitbold, E., Schraut, W., Moser, A. et al. Phase I study of a MUC1 vaccine composed of different doses of MUC1 peptide with SB-AS2 adjuvant in resected and locally advanced pancreatic cancer. Cancer Immunol Immunother 2005; 54: 254–264.

36. Rong, Y., Qin, X., Jin, D., Lou, W., Wu, L., Wang, D. et al. A phase I pilot trial of MUC1-peptidepulsed dendritic cells in the treatment of advanced pancreatic cancer. Clin Exp Med 2012; 12: 173–180.

37. Oki, Y. and Younes, A. Heat shock protein based cancer vaccines. Expert Rev Vaccines 2004; 3: 403–411.

38. Maki, R., Livingston, P., Lewis, J., Janetzki, S., Klimstra, D., Desantis, D. et al. A phase I pilot study of autologous heat shock protein vaccine HSPPC-96 in patients with resected pancreatic adenocarcinoma. Dig Dis Sci 2007; 52: 1964–1972.

39. Schmitz-Winnenthal FH, Grenacher L, Friedrich T, Lubenau H, Springer M, Breiner KM, et al. VXM01, an oral T-cell vaccine targeting the tumor vasculature: Results from a randomized,

controlled, first-in-man study in pancreatic cancer patients. J Clin Oncol 2103; 31(Suppl.): Abstract #3090.

40. Bae J, Carrasco R, Daley J, Dranoff G, Anderson KC, Munshi NC. Novel heteroclitic XBP1 peptides evoking antigen-specific cytotoxic T lymphocytes targeting various solid tumors. J Clin Oncol 2103; 31(Suppl.): Abstract #3067.

41. Chai K, Ai YQ, Jiang TW. Phase II study of dendritic cell vaccination combined with recombinant adenovirus-p53 in treatment for patients with advanced pancreatic carcinoma. J Clin Oncol 2103; 31(Suppl.): Abstract #3049.

42. Rossi GR, Hardacre JM, Mulcahy MF, Talamonti MS, Obel JC, Rocha Lima CMS, et al. Effect of algenpantucel-L immunotherapy for pancreatic cancer on anti-mesothelin antibody (Ab) titers and correlation with improved overall survival. J Clin Oncol 2103; 31(Suppl.): Abstract #3007.

43. Le DT, Wang-Gillam A, Picozzi VJ, Greten TF, Crocenzi TS, Springett GM, et al. Interim safety and efficacy analysis of a phase II, randomized study of GVAX pancreas and CRS-207 immunotherapy in patients with metastatic pancreatic cancer. J Clin Oncol 2103; 31(Suppl.): Abstract #4040.

44. Suzuki E, Kapoor V, Jassar AS, Kaiser LR, Albelda SM. Gemcitabine selectively eliminates splenic gr-1+/CD11b+ myeloid suppressor cells in tumor-bearing animals and enhances antitumor immune activity. Clin Cancer Res. 2005; 11(18):6713-6721.

45. Ghansah T, Vohra N, Kinney K, et al. Dendritic cell immunotherapy combined with gemcitabine chemotherapy enhances survival in a murine model of pancreatic carcinoma. Cancer Immunol Immunother. 2013; 62(6):1083-1091. doi: 10.1007/s00262-013-1407-9; 10.1007/s00262-013-1407-9.

46. Plate JM, Plate AE, Shott S, Bograd S, Harris JE. Effect of gemcitabine on immune cells in subjects with adenocarcinoma of the pancreas. Cancer Immunol Immunother. 2005; 54(9):915-925.

47. Shiao SL, Ganesan AP, Rugo HS, Coussens LM. Immune microenvironments in solid tumors: New targets for therapy. Genes Dev. 2011; 25(24):2559-2572.

48. Hassan R, Ho M. Mesothelin targeted cancer immunotherapy. Eur J Cancer. 2008 Jan; 44(1):46-53.

49. Le DT, Brockstedt DG, Nir-Paz R, et al. A live-attenuated Listeria vaccine (ANZ-100) and a live-attenuated Listeria vaccine expressing mesothelin (CRS-207) for advanced cancers: phase I studies of safety and immune induction. Clin Cancer Res. 2012 Feb 1;18(3):858-68

50. Rettig L, Seidenberg S, Parvanova I, Samaras P, Curioni A, Knuth A, Pascolo S. Gemcitabine depletes regulatory T-cells in human and mice and enhances triggering of vaccine-specific cytotoxic T-cells. Int J Cancer. 2011 Aug 15;129(4):832-8.

51. Williams S,C.P. No end in sight for telomerase-targeted cancer drugs. Nature Medicine 2013; **19**,6

52. Adams, S. Toll-like receptor agonists in cancer therapy Immunotherapy. 2009 November 1; 1(6): 949–964.

53. Royal RE, Levy C, Turner K et al. Phase 2 trial of single agent Ipilimumab (anti-CTLA-4) for locally advanced or metastatic pancreatic adenocarcinoma. J Immunother. 2010 Oct; 33(8):828-33.

54. Otahal P, Knowles BB, Tevethia SS, Schell TD. Anti-CD40 conditioning enhances the T(CD8) response to a highly tolerogenic epitope and subsequent immunotherapy of simian virus 40 T antigen-induced pancreatic tumors. J Immunol. 2007 Nov 15; 179(10):6686-95.

55. Khong A, Nelson DJ, Nowak AK, Lake RA, Robinson BW. The use of agonistic anti-CD40 therapy in treatments for cancer. Int Rev Immunol. 2012 Aug; 31(4):246-66.

56. Meili Zhang, Zhengsheng Yao, Sigrid Dubois, Interleukin-15 combined with an anti-CD40 antibody provides enhanced therapeutic efficacy for murine models of colon cancer PNAS 2009; 106 (18) 7513-7518

57. Jedd D. Wolchok, M.D., Ph.D., Harriet Kluger, M.D., et al. Nivolumab plus ipilimumab in advanced melanoma. N Engl J Med. 2013 Jun 2.